The Role of Smc3 in Mouse Embryonic and Adult Hematopoiesis

Tianjiao Wang
Washington University in St. Louis

Follow this and additional works at: https://openscholarship.wustl.edu/art_sci_etds

Part of the Cell Biology Commons

Recommended Citation
Wang, Tianjiao, "The Role of Smc3 in Mouse Embryonic and Adult Hematopoiesis" (2019). Arts & Sciences Electronic Theses and Dissertations. 1862.
https://openscholarship.wustl.edu/art_sci_etds/1862

This Dissertation is brought to you for free and open access by the Arts & Sciences at Washington University Open Scholarship. It has been accepted for inclusion in Arts & Sciences Electronic Theses and Dissertations by an authorized administrator of Washington University Open Scholarship. For more information, please contact digital@wumail.wustl.edu.
The Role of Smc3 in Mouse Embryonic and Adult Hematopoiesis

by

Tianjiao Wang

A dissertation presented to
The Graduate School of Washington University in partial fulfillment of the requirements for the degree of Doctor of Philosophy

May 2019
St. Louis, Missouri
Table of Contents

List of Figures ................................................................................................................................. iv

List of Tables ................................................................................................................................. vi

Acknowledgements ....................................................................................................................... vii

Abstract of the Dissertation ........................................................................................................... xi

Chapter 1: Introduction ..................................................................................................................... 1

1.1. Acute Myeloid Leukemia ......................................................................................................... 2

1.2 Cohesin in Cancer ...................................................................................................................... 16

Figure Legends ............................................................................................................................... 25

Figures ........................................................................................................................................... 26

Tables ............................................................................................................................................ 28

References ....................................................................................................................................... 30

Chapter 2: Smc3 is required for mouse embryonic and adult hematopoiesis................................. 42

Abstract .......................................................................................................................................... 43

Introduction ..................................................................................................................................... 43

Methods ......................................................................................................................................... 45

Results .......................................................................................................................................... 50

Discussion ...................................................................................................................................... 56

Figure Legends ............................................................................................................................... 62

Figures .......................................................................................................................................... 67

References ...................................................................................................................................... 79
Chapter 3: Exome analysis of treatment-related AML after APL suggests secondary evolution................................................................. 84

Figure Legends .................................................................................................................................................................................. 90

Figures ............................................................................................................................................................................................ 91

Tables ............................................................................................................................................................................................ 92

References ........................................................................................................................................................................................ 94
List of Figures

Chapter 1

Figure 1.1. Eight functional categories of genes that are frequently mutated in AML ....... 26

Figure 1.2. The cohesin complex .................................................................................... 27

Chapter 2

Figure 2.1. Generation of Smc3 conditional deficient mice and allele validation ......... 67

Figure 2.2. Embryonic hematopoietic Smc3 deletion ....................................................... 68

Figure 2.3. Homozygous somatic Smc3 deletion ................................................................. 69

Figure 2.4. Hematopoietic Smc3 haploinsufficiency .......................................................... 70

Figure 2.5. Competitive transplantation of Smc3 haploinsufficient bone marrow cells ..... 71

Figure 2.6. Effect of Dnmt3a haploinsufficiency on competitive disadvantage in Smc3 haploinsufficient BM cells ........................................................................................................ 72

Figure 2.7. Splicing analysis of exon 3 to exon 5 in wild-type and Smc3^{fl/+}/Vav1-Cre^{+/−} KL cells ......................................................................................................................... 73

Figure 2.8. Representative plot of intracellular flow cytometry data (Figure 2.1.D-E) ..... 74

Figure 2.9. Analysis of homozygous somatic Smc3 deletion ........................................... 75

Figure 2.10. Analysis of germline heterozygous Smc3 deletion ....................................... 76

Figure 2.11. Immunophenotypic analysis of colonies in serial replating assay ex vivo (Figure 2.4.C-E) ......................................................................................................................... 77

Figure 2.12 Competitive transplantation of ERT2-Cre^{+/−} bone marrow cells ............... 78
Chapter 3

Figure 3.1. Exome analysis of patient 10DD-1029 ................................................................. 91
List of Tables

Chapter 1

Table 1.1. Core subunits and regulatory proteins of the cohesin complex.............................. 28
Table 1.2. Studies of cohesin mutations in hematopoiesis......................................................... 29

Chapter 2

Chapter 3

Table 3.1. Clinical data of the patient ......................................................................................... 92
Table 3.2. APL patients with cytogenetic abnormalities in chromosomes 5 or 7 and evaluable subclonal architecture ........................................................................................................... 93
Acknowledgements

First and foremost, I would like to thank my thesis advisor, Dr. John Welch. This research could not have been completed without his continued guidance, support, and patience. I learned so much from John’s enthusiasm and dedication to improving treatment options for leukemia patients. He has been extremely encouraging over the years, especially in the last year of my graduate studies when I encountered difficulties. Thank you, John for believing in me when I had doubts and for cheering me all the way to the finish line.

I would like to express my gratitude to my wonderful thesis committee, Dr. Timothy Ley, Dr. Matthew Walter, Dr. Katherine Weilbeacher, Dr. Grant Challen. Tim is the best chair anyone could ask for. Thank you, Tim for all the great scientific discussion, kind advice, and for always keeping us on track. I rotated with Matt and began my journey on the floor in his lab. Thank you, Matt for introducing me to hematopoietic malignancies research and for providing insights whenever I have questions. Also, thank you for letting me use your printer all these years and for inviting me to your lab potlucks, cake sessions, and BBQ. Kathy was a huge support when I got scooped. Thank you, Kathy for teaching me to treat the incidence as a learning experience and for bringing your delightful enthusiasm for science to every committee meeting. I often run into Grant either in the mouse house or on my way to the mouse house. Thank you, Grant for being a role model of working hard and for giving me helpful suggestions on experiments.

Many thanks to members of the Welch lab. Thank you, Brandi for helping me with the project and for becoming my close friend. I miss the time we spend in and outside of lab together. Thank you for letting me be a part of Bradin’s life. It was a surreal experience welcoming him into the
world. Thank you, Orsola, my Italian sister. I wish you had come to the lab earlier and we would have more time together. It was a true pleasure getting to know you and being your friend. I love our coffee chats. Thank you for shining on me when the days were dark and for always caring about me. Thank you, Gayla for keeping our lab organized and for helping me with cloning Smc3. I have learned so much from your work ethic and your superb note-keeping skills. Thank you, Debbie and Conner for taking care of the mice over the years. I would like to also thank my past students Marti and David for the opportunity to mentor and learn from them.

I have been so lucky to be in a nurturing environment for my thesis research. The 6th and 7th floor of the Southwest tower is such a collaborative community. I want to thank Nikki for helping me with the cDNA library prep, Drs. Dave Spencer and Chris Miller for analyzing sequencing data, Julie for injecting mice for all my transplant experiments. I thank Bill, Dan, Matt at the Siteman Flow Cytometry Core for their expertise and help with my project. I am grateful for the instrumental support and guidance provided by Stacy and the MCB directors Drs. Jason Weber and Heather True. I am also thankful for the Siteman Cancer Center Cancer Biology Pathway funding that helped to make this work possible.

I would like to thank all the friends I have met during graduate school. Special thanks to Peter for being my person, whom I can always talk to and count on, and for going through the PhD years together. I also thank Brian, Ling, Tanzir, Monique, KK, Cara, Neal, Hamza for help with my experiments, words of wisdom, and great memories. Thank you to my friends from the entering class of 2012 - Kyle, Linxuan, Bhavna, Melissa, Sarem, Terin for making this journey so memorable.
I would like to thank my families away from home, the Capozzola’s and the Serlin’s. Thank you, Kate, Shannon, Mr & Mrs. Capozzola, Grandma and Grandpa Toomey for taking me in as your sister, daughter, and granddaughter and loving me for the past 10 years. Thank you, Robert for all the wonderful conversations and career help. Guillemette, Felix, Lukas, Navit, and Marec, I love all of you guys. Words cannot express my gratitude to my dear friends, Mo, Jeff, Fujun, Sicong, Huakang, and Qianwen. Thank you for the late nights, early mornings, and the countless messages. Thank you for all your love and support.

Finally, I would like to thank my family. Heartfelt thanks to my grandparents for being my biggest cheerleaders and for giving me strength. Thank you, Dad for the witty comfort and the unconditional support. Thank you, Mom for all the sacrifices you have made to help me achieve my goals, for taking care of me and keeping me company. Thank you, Dumpling, my dear cat for finding me on the street 2 years ago and following me home. You have been the source of my happiness. Thank you for always being with me.

Tianjiao Jephne Wang

Washington University in St. Louis

May 2019
Dedicated to my parents and grandparents

This work is impossible without your love and unwavering support

And to my beloved cat, Dumpling

Who is always with me, for better, or for worse
Acute myeloid leukemia (AML) is a heterogeneous disease, characterized by recurrent genetic mutations. Mutations in the cohesin complex are one of the 8 functional categories of mutations in AML. *SMC3* encodes a subunit of the cohesin complex, which has important roles in chromosome segregation, genome instability, and gene expression. In the first chapter of the dissertation, we discuss the genetics of AML, normal functions of the cohesin complex, and the interplay between cohesin mutations and myeloid malignancies.

*SMC3* is recurrently mutated in AML and other myeloid malignancies. In the second chapter of the dissertation, we compare the consequences of *Smc3* deficient and haploinsufficient mouse models to determine whether the heterozygous missense mutations in *SMC3* might have dominant-negative effects or phenocopy loss-of-function effects. We found that homozygous deletion of *Smc3* during embryogenesis or in adult mice resulted in hematopoietic failure. *SMC3* missense mutations are therefore unlikely to be associated with simple dominant negative phenotypes due to incompatibility with hematopoiesis. *Smc3* haploinsufficiency, in contrast, was
tolerated during embryonic and adult hematopoiesis. Under steady-state conditions, Smc3 haploinsufficiency did not alter colony forming capacity ex vivo and led to modest transcriptional and chromatin accessibility changes in Lin-cKit+ progenitor cells. However, following tamoxifen-induced deletion in competitive transplantations, we observed a significant hematopoietic competitive disadvantage in Smc3 haploinsufficient bone marrow cells across myeloid and lymphoid lineages and within the stem/progenitor compartments. The competitive disadvantage was not affected by different conditions of hematopoietic stresses, but was partially abrogated by concurrent Dnmt3a haploinsufficiency, suggesting that antecedent mutations may be the prerequisites to realize the leukemogenic potential of Smc3 mutations.

In the third chapter of the dissertation, we present a case of an older women that initially appeared to be treatment-related AML following non-cytotoxic all-trans retinoic acid (ATRA)/arsenic trioxide (ATO) therapy for acute promyelocytic leukemia (APL), but upon further analysis found to be more consistent with secondary AML. Exome sequencing revealed a TET2-mutated dominant clonal process that preceded the APL diagnosis, persisted, and gave rise to an AML-associated new subclone with a NPM1 mutation. Review of additional cytogenetic abnormalities observed in APL patients showed that cytogenetic abnormalities commonly occur as subclones of the APL clone, although one rare case with del(7) independent of the APL clone was identified. These results demonstrated that APL may emerge within the context of clonal hematopoiesis and caution must be exercised when interpreting the development of tAML after ATRA/ATO therapy, especially in older patients.
Chapter 1:

Introduction
1.1 Acute Myeloid Leukemia (AML)

1.1.1 Disease statistics

Acute myeloid leukemia is an aggressive myeloid neoplasm characterized by accumulation of myeloblasts in the blood or bone marrow.\(^1\) Proliferating immature myeloblasts impair the development of normal hematopoiesis, leading to severe infections, cytopenias, anemia, immune compromise, and death.\(^2\) AML is the most common acute leukemia in adults, with 19,520 estimated new cases in 2018, accounting for 1.1% of all new cancer cases in the US.\(^3\),\(^4\) AML is slightly more common among men than women, and approximately 0.5% of the population will be affected at some point during their lifetime based on 2013-2015 data.\(^4\),\(^5\) Although AML can occur in any age group, AML is primarily a disease of the elderly, with a median age at diagnosis of 68 years.\(^5\),\(^6\) Advances in the treatment of AML have significantly improved the outcomes for younger adult patients, with 5-year survivals of 35 to 40% among those who are 60 years of age or younger.\(^7\) However, prognosis in older patients, who account for the majority of new cases, remains dismal, with 2-year survivals of only 5 to 15% among patients who are older than 60 years of age, as much as 70% of the elderly will die within 1 year of diagnosis.\(^7\),\(^8\) Across all age groups, the 5-year overall survival of AML is 27.4%, with an estimated 10,670 deaths in 2018, consisting of 1.8% of all cancer deaths in the US.\(^4\)

1.1.2 Genetics

1.1.2.1 AML with recurrent genetic abnormalities

AML is a heterogeneous disease. The cytogenetic heterogeneity of AML has been recognized for more than three decades. Based on karyotype analysis, AML with recurrent genetic
abnormalities can be divided into two subtypes: (1) AML with chromosomal aneuploidies; (2) AML with balanced genomic rearrangements.  

**AML with chromosomal aneuploidies**

Over 60% of cases in the subgroup of AML with chromosomal aneuploidies have at least 3 chromosomal events, of which the most frequent are -5/5q, -7/7q, -12/12p, -17/17p, and +8/8q. Approximately 50% of patients with deletions in chromosomes 5, 7, 12, or 17 have *TP53* mutations, and these are more commonly observed in older patients; the median age of patients with chromosomal aneuploidy and *TP53* mutations is 58 years vs. 49 years with aneuploidy alone. Patients with both complex karyotype and *TP53* mutations have significantly inferior prognosis to the poor overall outcomes associated with either subset alone, but recent data by groups at Washington University in St. Louis suggest that this unique subgroup of AML may respond favorably to hypomethylating agent, decitabine.

**AML with balanced genomic rearrangements**

AML with balanced genomic rearrangements tend to present at a younger age and have, on average, 1 genomic rearrangement and lower overall number of acquired mutations, most frequently concurrent with activating mutations *FLT3*-internal tandem duplication (ITD), *KIT*, *NRAS*, tyrosine or serine-threonine kinases, and protein tyrosine phosphatases. There are at least 7 distinct subtypes of recurrent genomic rearrangements in AML, each defining a clinicopathologic entity. These translocations and inversions are considered leukemia-initiating and are almost uniformly present in patients who subsequently relapse.

The most common translocation fusion gene is *PML-RARA*, defined by t(15;17)(q22;q21), which occurs in 5-13% of patients and is characteristic of acute promyelocytic leukemia (APL). *FLT3-*
ITD and WT1 mutations co-occur with PML-RARA in approximately 35% and 15% of APL cases, respectively. APL patients with PML-RARA that are FLT3 negative are associated with favorable outcomes when treated with combinational chemotherapy that includes all-trans-retinoic acid (ATRA).\textsuperscript{13} Outcomes in APL patients treated with chemotherapy alone were historically dismal, demonstrating the adaptive relevance of mutation: treatment interactions.

**RUNX1-RUNXIT1** AML, defined by t(8;21)(q22;q22.1), occurs in 1-6% of patients, is associated with good risk following treatment with high dose cytarabine. KIT mutations co-occur with t(8;21) in approximately 25% of **RUNX1-RUNXIT1** AML, and these patients have inferior outcomes compared to KIT wild type patients.\textsuperscript{13, 14}

The **CBFB-MYH11** fusion results from inv(16)(p13.1q22) and occurs in 1-6% of AML patients. **CBFB-MYH11** AML also has favorable prognosis in the absence of KIT mutations. NRAS mutations co-occur with **CBFB-MYH11** in approximately 40% of AML cases. The less frequent genomic rearrangements, affecting about 1% or less of AML patients include: **MLLT3-KMT2A**, defined by t (9;11)(p21.3;q23.3); **DEK-NUP214**, defined by t(6;9)(p23;q34.1); GATA2, **MECOM**, defined by inv(3)(q21.3q26.2), and **RBM15-MKL1**, defined by (t1;22)(p13.3;q13.3).\textsuperscript{9}

1.1.2.2 AML with gene mutations

Over the past 15 years, advances in next-generation sequencing (NGS) have tremendously increased our knowledge of the molecular heterogeneity of AML. AML was the first primary cancer to be studied by massively-parallel sequencing technologies.\textsuperscript{15} In 2008, the first AML genome was published in a landmark study done by groups at Washington University in St. Louis.\textsuperscript{16} Subsequent studies have identified numerous novel recurrent somatic mutations with
biologic, prognostic, and therapeutic relevance and have demonstrated that AML is a complex and dynamic disease. Emerging data with the use of NGS are revolutionizing our view of the spectrum and frequency of mutations, their distinct patterns of cooperativity and mutual exclusivity, their subclonal architecture, the epigenetic landscape of the disease, and the clonal evolution during AML.\textsuperscript{17}

\textit{Clonal Evolution}

Studies have shown that most cases of AML are characterized by clonal heterogeneity at the time of diagnosis, with more than half of the patients exhibiting at least one subclone in addition to a founding clone.\textsuperscript{11} Data from clonal evolution studies provide support for a model that mutations in genes involved in epigenetic regulation (specifically genes involved in the regulation of DNA methylation and chromatin modifications, most commonly \textit{DNMT3A}, \textit{TET2}, \textit{IDH1}, \textit{IDH2}, and \textit{ASXL1}) are present in preleukemic hematopoietic stem cells (HSCs) and occur early in the evolution of AML, preceding secondary leukemogenic events such as mutations in nucleophosmin (\textit{NPM1}) or signaling genes (\textit{FLT3}, \textit{RAS}).\textsuperscript{13, 17} Furthermore, the epigenetic modifying genes are frequently found to be mutated in elderly individuals along with clonal expansion of hematopoiesis that confers and increased risk for the development of hematologic cancers.\textsuperscript{13} Such ancestral preleukemic stem cells are capable of multilineage differentiation. For example, preleukemic \textit{DNMT3A}-mutant HSCs were shown to have a multilineage repopulation advantage over wild type HSCs and were detected in samples collected from patients who were in morphologic complete remission, indicating their potential to be resistant and survive chemotherapy.\textsuperscript{18} Thus, preleukemic hematopoietic clones can persist over time, survive chemotherapy, expand during remission, and eventually leading to relapse and the various patterns of clonal composition that occur at relapse may contribute to resistance to therapy.\textsuperscript{19}
**Clonal Hematopoiesis**

Recent studies of large population-based cohorts show that clonal hematopoiesis with recurrent mutations in epigenetic regulators *DNMT3A*, *TET2*, and *ASXL1* (and less frequently in splicing factor genes *SRSF2*, *SF3B1* and in the genotoxic sensor *TP53*) increases as people age and confers an increased risk of hematologic cancer and death.\(^{20, 21, 22, 23, 24}\) Expanded clones containing these somatic mutations can be identified in the blood or bone marrow of patients without evidence of overt hematologic malignancy and decades before the development of AML. This defines a new entity, termed either “age-related clonal hematopoiesis” (ARCH) or “clonal hematopoiesis of indeterminate potential (CHIP)”, which has been identified in approximately 10% of patients 70-80 years old.\(^{22, 23}\) A recent study conducted by groups at Washington University in St. Louis using bar-coded sequencing found a higher incidence of ARCH if the threshold of detection is lowered to 0.5%.\(^{25}\) The incidence of CHIP increases with age, predisposes patients to AML and other hematologic malignancies, including myelodysplastic syndromes (MDS), and the transformation rate of CHIP into a hematologic disease is about 0.5-1% per year.\(^{26}\)

**The Cancer Genome Atlas (TCGA) Project**

The Cancer Genome Atlas Research Network analyzed the mutational profiling of 200 patients with de novo AML by either whole-genome (n=50) or whole-exome (n=150) sequencing, along with RNA and microRNA expression and DNA methylation analysis.\(^{11}\) Significantly mutated genes in AML were organized into 8 functional categories, summarized in Figure 1.1.\(^{17}\) (1) Mutations in *NPM1*, encoding a multifunctional nucleo-cytoplasmic shuttling protein, resulting in the aberrant cytoplasmic localization of NPM1 and NPM1-interacting proteins; (2) Mutations in signaling genes such as kinases *FLT3*, *KIT*, or RAS family members *KRAS*, *NRAS* that confer
a proliferative advantage through the RAS-RAF, JAK-STAT, and PI3K-AKT signaling pathways; (3) Mutations in myeloid transcription factors such as RUNX1 and CEBPA, leading to transcriptional deregulation and impaired hematopoietic differentiation; (4) Mutations in tumor-suppressor genes such as TP53 and WT1 that result in transcriptional deregulation and impaired degradation through the mouse double minute 2 homologue (MDM2) and the phosphatase and tensin homologue (PTEN); (5) Mutations in DNA methylation-associated genes DNMT3A and TET2 that deregulate DNA methylation patterns and lead to transcriptional deregulation of leukemia-associated gens or in IDH1 and IDH2 that act through the 2-hydroxyglutarate (2-HG) oncometabolite production and impact DNA methylation via impairment of TET2; (6) Mutations in chromatin-modifying genes such as AXL1 and PHF6, leading to deregulation of chromatin modification, for instance methylation of histones H3 and H2A; (7) Mutations in spliceosome-complex genes such as SRSF2, SF3B1, U2AF1, and ZRSR2 that are involved in impaired spliceosome function and deregulated RNA processing; (8) Mutations in cohesin-complex genes such as SMC3, STAG2, and RAD21 that may impair accurate chromosome segregation and transcriptional regulation.17

**NPM1 mutations**

*NPM1* encodes a phosphoprotein that normally shuttles between the nucleus and the cytoplasm and plays a role in in epigenetic control, ribosomal protein assembly, and regulation of p53 tumor suppressor pathway.27 *NPM1* mutations are the most common genetic mutations in AML, found in approximately 30% of all AML and 45-60% of AML with normal karyotype.17,28 They are mutually exclusive to other genomic rearrangements and frequently co-exist with DNMT3A (approximately 50%), FLT3-ITD (approximately 40%), NRAS (approximately 20%), cohesin genes SMC3, SMC1A, RAD21 (approximately 20%), TET2 (approximately 15%), IDH1
Mutations in DNA hydroxymethylation genes (DNMT3A, TET2, IDH1, and IDH2) typically represent the first acquired event and are present in the founding clone while NPM1 is acquired as a secondary event during leukemogenesis, together with mutations in FLT3, NRAS, and PTPN11. In younger patients (<60 years old), NPM1 mutations in cytogenetically normal AML without FLT3-ITD mutations portend a favorable prognosis. However, patients with concomitant mutations in NPM1, FLT3-ITD, and DNMT3A, which represent the most frequent triple genotype in AML, have significantly shorter event-free survival and inferior overall survival.

**Mutations in signaling genes**

FLT3 encodes a receptor tyrosine kinase involved in hematopoiesis. There are two common mutations that occur in FLT3: ITD in the juxtamembrane domain and a point mutation of the tyrosine kinase domain (TKD), both mutations lead to constitutive activation. Approximately 20% of all AML cases harbor a FLT3-ITD mutation, which is associated with an unfavorable prognosis and the mutation is more common in cytogenetically normal AML, accounting for approximately 30% of these cases. The frequency of FLT3-ITD mutations decreases with older age and FLT3 mutations are associated with NPM1 mutations. There is variability in the size of the FLT3-ITD, ranging from a few base pairs to over 1000 base pairs, the number of FLT3-ITD mutations, approximately 14-25% of FLT3-ITD positive patients will have more than one FLT3-ITD mutation. Sequencing of FLT3-ITD reveals that the sequence and site of the mutations are variable: in fact, only about two-thirds of the FLT3 mutations are true tandem duplications while the remaining are insertions or complex duplications and insertions; approximately 30% of FLT3-ITD occur outside the juxtamembrane domain and instead occur in the TKD, usually in the
β1 sheet. The less common FLT3-TKD mutation is found in approximately 10% of AML.

*KIT* encodes a receptor tyrosine kinase that plays important roles in proliferation, differentiation, and cell survival. The ligand for KIT is stem cell factor (SCF). Binding of SCF to the extracellular domain of KIT induces receptor dimerization and activation of downstream signaling pathways that are involved in mediating pro-growth and pro-survival signals within the cell, including the MAPK signaling pathway (RAS-RAF-MEK-ERK), the PI3K pathway (PI3K-AKT-mTOR), and the STAT3 pathway. KIT mutations are gain-of-function mutations that occur in less than 5% of all AML cases and are higher, 25-35% of cases in core-binding factor leukemia. KIT mutations occur primarily in exon 17 and affect the activation loop of the kinase domain, resulting in improved proliferation and survival of leukemic cells. KIT mutations confer unfavorable prognosis in AML with t(8;21), RUNX1-RUNXIT1 AML.

*KRAS* and *NRAS* belong to the RAS GTPase family of genes. *KRAS* mutations are less common in adults, found in only 2% of cases vs. 9% of cases in children. NRAS mutations occur in approximately 15% of AML cases in adults and children. The concurrent mutations of NRAS are NPM1 and biallelic CEPBA. RAS mutations do not appear to have a clear impact on outcome except for NRAS*G12/G13*, which confers superior outcomes in presence of NPM1 and DNMT3A mutations.

*Mutations in myeloid transcription factors*

*RUNXI* encodes the alpha subunit of the heterodimer core binding factor, which is involved in transcription. Somatic *RUNXI* mutations occur in 5-20% of AML and the incidence increases with older age. They co-segregate with mutations in SRSF2 (approximately 25%), ASXL1
(approximately 20%), KMT2A (15-20%), IDH2R140 (approximately 12%). They are mutually exclusive with NPM1, biallelic CEBPA, and AML with recurrent cytogenetic abnormalities. RUNX1 mutations are associated with male sex, inferior outcome, and secondary AML evolving from MDS. Germline RUNX1 mutations are found in the autosomal dominant familial platelet disorder, conferring a predisposition to AML.

CEBPA encodes a transcription factor involved in granulocytes differentiation. CEBPA mutations are found in approximately 10% of AML and are more common in cytogenetically normal AML or with 9q deletions. The incidence of CEBPA mutations declines with older age. Approximately 2/3 of CEBPA mutations may be biallelic, which usually include one N-terminus and one C-terminus mutation, leading to null expression of CEBPA, and the rest are monoallelic, which can be truncating N-terminal mutations resulting in a shortened CEBPA with a dominant negative effect or C-terminal mutations that decrease dimerization or DNA binding.

Biallelic CEBPA mutations co-occur with NRAS (approximately 30%), GATA2 (approximately 30%), WT1 (approximately 20%), CSF3R (approximately 20%), and 9q- (approximately 15%), and confer a favorable prognosis.

Mutations in tumor-suppressor genes

TP53 is a tumor suppressor gene and frequently referred to as the “guardian of the genome” that regulates the cell cycle in response to cellular stresses. TP53 mutations occur in 5-20% of adult AML and approximately 1% of pediatric AML. The incidence of TP53 mutations significantly increases with older age. TP53 mutations are predominantly detected in AML with complex karyotype (56-78% of cases) and are associated with very poor outcome in AML as in other cancers.
WT1 encodes a transcription factor important for normal cellular development and cell survival that appears to play a tumor suppressor role in renal tissues, but an oncogenic role in leukemia. WT1 mutations can be found in 4-11% of AML cases and are linked with poor outcome in AML with a normal karyotype.

Mutations in DNA-methylation-associated genes

DNMT3A encodes a DNA methyltransferase involved in the epigenetic regulation of the genome through methylation. DNMT3A mutations are quite common in AML, occurring in approximately 20% of patients and frequently co-occur with NPM1, FLT3-ITD, IDH1, IDH2, and SMC3 mutations.17,49 The most common mutation is a substitution of arginine at position 882 (R882).50 DNMT3A with heterozygous R882H mutation forms stable heterodimers with wild type DNMT3A, disrupting the ability of the wild type DNMT3A protein to form active tetrarimers and leading to a hypomorphic effect on the methyltransferase activity of the enzyme and also a dominant negative effect on the wild type DNMT3A.51,52,53 The incidence of DNMT3A mutations increases with older age. They are associated with CHIP and secondary AML evolving from MDS and are early events in leukemogenesis. The frequency of DNMT3A R882 mutations is less than one-third of CHIP DNMT3A mutations, but more than two-thirds of AML DNMT3A mutations. DNMT3A mutations have moderate adverse effect on outcome, which can be overcome by high doses anthracycline chemotherapy.47,38

TET2 encodes an epigenetic modifier that converts methylcytosine to 5-hydroxymethylcytosine and is also involved in myelopoiesis. TET2 mutations are found in 7-25% of adult AML and 1.5-4% of pediatric AML and are early events in leukemogenesis. Mutations in TET2 are highly
variable, including nonsense mutations, deletions, missense mutations, and splice-site mutations, which all appear to cause loss-of-function and decrease hydroxymethylation of DNA.\textsuperscript{54} NPM1 mutations and TET2 mutations statistically co-occur with FLT3-ITD and -TKD aberrations.\textsuperscript{55} In contrast, IDH mutations seldom co-exist with TET2 mutations possibly because 2-HG inhibits the activity of TET2 (see below).\textsuperscript{55,56} The incidence of TET2 mutations in AML increases with older age and TET2 mutations have been found in healthy elderly individuals with CHIP.\textsuperscript{20}

IDH1 and IDH2 are genes involved in metabolism and may also play an epigenetic role in histone and DNA methylation.\textsuperscript{57} Mutations in IDH1 and IDH2 occur at the active isocitrate binding site, which alters the enzymatic activity and leads to the generation of a novel oncometabolite, 2-HG.\textsuperscript{58} IDH mutations statistically co-occur with NPM1 mutations (except for IDH2\textsuperscript{R172}).\textsuperscript{59} They are associated with CHIP in healthy elderly individuals (although much less commonly than DNMT3A, TET2, ASXL1, and TP53 mutations) and are early events in leukemogenesis.\textsuperscript{13} IDH1 mutations affect the arginine at position 132 or 170 (R132 or R170) and can be found in 7-14\% of adult AML cases, but only 1\% of pediatric AML.\textsuperscript{17,37} These mutations are mutually exclusive and exclusive of the IDH2 mutations. IDH1 mutations are associated with unfavorable outcome.\textsuperscript{60} IDH2 mutations affect the arginine at position 140 or 172 (R140 or R172) and occur in 8-19\% of adult AML, but only 1-2\% of pediatric cases.\textsuperscript{17,61} The incidence of IDH2\textsuperscript{R140} mutation increases with older age and has been shown to have a favorable prognosis in intermediate risk AML with NPM1 mutations.\textsuperscript{38}

Mutations in chromatin-modifying genes

ASXL1 encodes a chromatin binding protein, which regulates gene transcription in localized areas via modifying chromatin structure. ASXL1 mutations are frequently found in MDS and AML, with a frequency of 5-15\%, but appear to be enriched in secondary AML and intermediate
risk AML. ASXL1 mutations are associated with male sex and CHIP in healthy elderly people and they also increase with older age, more prevalent in patients over 60 years old and quite rare (approximately 1%) in children. Frequent concomitant mutations are RUNX1 (approximately 20%), IDH2R140 (approximately 13%), and SRSF2. ASXL1 mutations are early events in leukemogenesis, with most studies showing they are predictive of inferior outcome, particularly genotypes ASXL1mut/RUNX1mut and ASXL1mut/SRSF2mut.17

PHF6 is an X-linked gene that appears to be a highly dynamic chromatin adaptor protein that interacts with a growing number of partners (nucleosome remodeling and deacetylation complex, PAF1, UBF) to regulate transcription.64 Germline loss-of-function mutations in PHF6 are the cause of the Börjeson-Forssman-Lehmann X-linked intellectual disability syndrome.65 Somatic PHF6 mutations occur in 2-3% of adult AML and are more frequent in males than females.66, 67 They are associated with adverse prognosis in intermediate risk AML patients who are negative for FLT3-ITD.38

Mutations in spliceosome-complex genes

Mutations in splicing factor genes SRSF2, SF3B1, U2AF1, and ZRSR2 lead to impaired spliceosome function and deregulated RNA processing resulting in aberrant splicing patterns. Mutations in spliceosome-complex genes account for 14% of AML patients in the TCGA cohort.11 They are associated with CHIP in healthy elderly persons and poor outcome, shown by a few studies on clinical significance.13 Moreover, mutations of splicing factors occur in high frequencies in MDS. Refractory anemia with ringed sideroblasts (RARS) is a subtype of MDS characterized by the accumulation of erythroid precursor cells and 15% or more ring sideroblasts in the bone marrow. SF3B1 is highly mutated in RARS, whereas U2AF1 mutations are not linked with ringed sideroblasts and RARS.68
SRSF2 mutations are also found in chronic myelomonocytic leukemia (CMML) and confer an increased risk of transformation to acute leukemia. Mutations can occur in multiple domains, although the most recurrent mutations affect the RNA recognition motif and arginine/serine-rich protein interaction domain of the protein.\textsuperscript{69,70} Functional studies have shown that the P95 SRSF2 mutations have an altered RNA-binding activity resulting in mis-splicing of many important genes including \textit{EZH2}.\textsuperscript{71}

\textit{SF3B1} is the most commonly mutated spliceosomal gene in hematological cancers, including MDS, with almost half of \textit{SF3B1} mutations in lysine 700. Heterozygous \textit{SF3B1} mutations are mostly missense substitutions in addition to hotspots in the HEAT repeat domains.\textsuperscript{71}

\textit{U2AF1} is frequently mutated in codons S34 and Q157 in approximately 11\% of MDS patients. Heterozygous insertions and deletions have also been reported.\textsuperscript{72,73} \textit{U2AF1} mutations appear to interfere with 3’ splice site binding function of the protein, leading to aberrant alternative splicing of numerous U2-dependent introns potentially and constructing an entirely novel transcriptome specific to MDS.\textsuperscript{74}

\textit{ZRSR2} mutations are distributed throughout the gene in MDS patients, interrupting the coding capacity by creating in-frame stop codons and therefore suggestive of loss-of-function phenotypes.\textsuperscript{72} Knockdown of \textit{ZRSR2} revealed a distinct splicing defect pattern of the U12-dependent introns, affecting a large number of U12-type intron-containing genes that play a significant role in MAPK signaling pathways and E2F transcription activities, and impaired \textit{in vitro} erythroid differentiation while promoted myeloid differentiation of cord blood-derived CD34+ cells, which supports MDS phenotype.\textsuperscript{75}

\textit{Mutations in cohesin-complex genes}
Mutations in cohesin complex genes \textit{SMC3}, \textit{STAG2}, \textit{RAD21}, and \textit{SMC1A} may cause defects in chromatid cohesion or impact transcriptional regulation. Cohesin mutations occurred in about 10\% of non-M3 AML cases and were identified in 13\% of AML patients in the TCGA cohort.\textsuperscript{11} Cohesin mutations frequently co-occur with \textit{NPM1} mutations and \textit{RUNXI-RUNXIT1}. Other common mutations concurrent with cohesin mutations in AML include \textit{RAS}, \textit{RUNX1}, \textit{TET2}, \textit{ASXL1}, and \textit{EZH2}.\textsuperscript{11,39,76} Cohesin mutations are not only found in AML, but also in other myeloid malignancies such as CMML, chronic myeloid leukemia (CML), myeloproliferative neoplasms (MPNs), and MDS.\textsuperscript{77,78} Notably, more than 50\% of patients with Down syndrome-associated acute megakaryocytic leukemia (DS-AMKL) have mutations in \textit{STAG2}, which can co-occur with mutations in \textit{RAS}, \textit{ASXL1}, \textit{EZH2}, \textit{JAK2}, and \textit{JAK3}.\textsuperscript{79}

\textit{SMC3} and \textit{RAD21} mutations are nearly universally heterozygous and the majority of \textit{SMC3} mutations are missense mutations. Intriguingly, \textit{SMC3} mutations frequently co-occur with \textit{DNMT3A} mutations, one of the most commonly mutated genes in AML. \textit{STAG2} and \textit{SMC1A} are encoded on the X chromosome, and therefore mutations would be thought to result in null alleles.\textsuperscript{80} Additionally, cohesin mutations tend to be mutually exclusive, implying that either they may not be tolerated by a cell when co-occurring or alteration in one component may be sufficient to disrupt the entire complex.\textsuperscript{49,76} Although cohesin mutations are often observed as early subclonal events during leukemia development, conceivably facilitating disease initiation, they are not observed in CHIP; thus, they are unlikely to be the initiating event.\textsuperscript{21,81,82} In most AML cases, cohesin mutations are not associated with karyotypic abnormalities, suggesting cohesin mutations contribute to leukemogenesis through alternative pathways other than inducing genomic instability.\textsuperscript{11,76}
This thesis focuses on understanding the contribution of cohesin mutations, particularly SMC3 mutations to the pathogenesis of AML.

1.2 Cohesin in Cancer

1.2.1 Roles of cohesin

The cohesin complex consists of four core subunits, structural maintenance of chromosomes (SMC) proteins SMC1A and SMC3, RAD21, and STAG. In mammals, there are two related STAG proteins, STAG1 or STAG2. Both SMC proteins are rod-shaped proteins containing ATP-binding cassette (ABC)-like ATPase motifs and are characterized by a globular hinge domain flanked by two alpha-helical domains, which fold back on themselves at the hinge, forming a long antiparallel coiled coil arm that brings the N- and C-termini together. SMC1A and SMC3 form a V-shaped heterodimer at the hinge domains. At the distal end of the two coiled coil arms, the N- and C-termini of each SMC protein form an ATPase head domain. The kleisin family protein RAD21 physically connects the ATPase heads of SMC1A and SMC3, thus forming a tripartite ring-like structure, with an internal diameter of about 40nm. The STAG subunit interacts with RAD21 and further stabilizes the cohesin ring. In addition to the four core subunits, cohesin loaders (Scc2/NIPBL, Scc4/MAU2), cohesin regulators (PDS5, SORORIN, and WAPL), cohesin protector (SGOL), and cohesin modifiers (ESCO and HDAC8) also bind to or modify the cohesin complex (Figure 1.2).

The cohesin complex is highly conserved through evolution with homologs in yeast, fruit flies, and mammals (Table 1.1). Among the several models have been proposed to depict how the cohesin complex associates with chromatin, the one-ring “embrace” model and the two-ring “handcuff” model are supported by experimental data. The one-ring model suggests that the
cohesin ring embraces two chromatins until their segregation. The two-ring model describes each cohesin ring entraps one chromatin and cohesion is mediated by interactions between the two cohesin rings. The canonical role of the cohesin complex is to ensure proper segregation of chromosomes during mitosis and meiosis. In addition to its essential role in sister chromatids cohesion, cohesin contributes to genome maintenance and functions by involving in DNA damage repair and gene expression.

1.2.1.1 Cohesin functions in chromosome segregation

Cohesins are loaded to the chromatins at the G1/S phase in yeast and at telophase in mammalian cells by loading complex NIPBL-MAU2. During DNA replication at S phase, each cohesin ring embraces one of the sister chromatids. After DNA replication at the S phase, acetylation of SMC3 by cohesin acetyltransferases ESCO1 and ESCO2 establishes stable cohesion between the newly replicated sister chromatids. PDS5 and SORORIN form a complex to maintain the cohesion throughout the G2 phase until prophase when SORORIN is phosphorylated and destabilized. The removal of cohesins are facilitated by the formation of PDS5-WAPL complex. At prophase, cohesins on the chromosomal arms are removed by the phosphorylation of RAD21 and STAG1/2 by PLK1. Centromeric cohesion is protected by SGOL1 until Separase gets activated and cleaves RAD21 at anaphase and therefore separating the sister chromatids. The dissociated cohesins can be recycled after the acetyl groups are removed from SMC3 proteins by cohesin deacetylases HDAC8. In meiosis, a similar biphasic removal of cohesin occurs, with RAD21 replaced by REC8.

The main roles of cohesin during cell cycle are to keep sister chromatids together and to provide resistance when sister chromatids are pulled by microtubules towards the opposing spindle poles, thus ensuring accurate separation of sister chromatids during the transition from metaphase to
anaphase. Failure in the formation and maintenance of sister chromatid cohesion results in premature chromosome segregation, which is thought to be a major pathway to aneuploidy, a characteristic observed in many human cancers.

1.2.1.2 Cohesin functions in genome instability

Cellular DNA is exposed to single and double strand breaks (DSBs) through multiple endogenous and exogenous mechanisms. Cells respond to DNA damage by activation of DNA-damage checkpoints that halt cell cycle progression until the damaged DNA is repaired. If the damage cannot be repaired properly, cells may undergo apoptosis. Eukaryote cells have two distinct mechanisms to repair DSBs, the homologous recombination (HR) between sister chromatids in the S and G2 phases and the non-homologous end joining (NHEJ), involving re-ligation of broken DNA, which occurs throughout the cell cycle.

The function of cohesin in DNA damage repair is evolutionarily conserved from yeast to humans. Rad21 was cloned originally by complementing the γ-radiation sensitivity in fission yeast with a function in DSB repair, before its role in sister chromatid cohesion was identified. In response to laser-induced DNA damage in human cells, cohesins are recruited to the DSB site and de novo cohesion, named damage-induced cohesion (DI-cohesion), is established. Besides cohesins, factors that are required to load cohesins to chromatin, establish cohesion, and maintain cohesion are all needed for DNA damage repair. Defects in the cohesin-loading complex NIPBL-MAU2, cohesin acetyltransferase ESCO, or maintenance factor SORORIN block the accumulation of cohesins at DSBs and prevent DNA damage repair, suggesting the presence of cohesins on chromatin is not sufficient to mediate DNA repair and instead, additional cohesion is required. DI-cohesion may help to structurally stabilize chromosomes whose DNA backbone has been fragmented by DSBs and to provide the proximity
between the damaged sister chromatid and the template, allowing HR to occur. DNA damage-
induced phosphorylation and acetylation on SMC3 were found to be important for genome-wide
DI-cohesion caused by DSB in the G2 phase and DSB repair.\textsuperscript{112}

Moreover, cohesins are required to activate checkpoints when DSBs occur.\textsuperscript{116} In \textit{C. elegans},
when SCC2, a component of the cohesin loading complex, is mutated, cohesins cannot be loaded
onto chromatin in meiosis, resulting in failure of both checkpoint activation and DNA damage
repair.\textsuperscript{117} This shows the importance of cohesin recruitment to the damaged chromatin. The
checkpoint role of cohesins is independent of its function in sister chromatid cohesion because
cohesins are required for the phosphorylation and activation of Chk2 although no sister
chromatid cohesion occurs in the G1 phase. As evidence, depletion of SORORIN, a protein
essential for the generation and maintenance of sister chromatid cohesion, leads to checkpoint
activation but DSB repair failure.\textsuperscript{116}

1.2.1.3 Cohesin functions in gene expression

The first evidence that cohesin factors regulate gene expression and development came from the
studies of \textit{Drosophila cut} and \textit{Ultrabithorax} genes: heterozygous Nipped-B mutants showed
reduced \textit{cut} expression, whereas loss of Smc1, Rad21, or SA led to increased \textit{cut} expression.\textsuperscript{118,119} Cohesins also facilitate expression of \textit{c-myc}, a function conserved across \textit{Drosophila},
zebrafish, mouse, and humans and cohesin depletion reduces \textit{myc} transcription.\textsuperscript{120} Furthermore,
cohesins present in non-cycling and even post-mitotic cell in higher eukaryotes.\textsuperscript{121} Accumulating
evidence implies an important non-canonical role of cohesin in regulating gene expression,
which is independent of cohesins’ role in cell division.\textsuperscript{83} In non-dividing mouse thymocytes,
genetic deletion of cohesin resulted in reduced transcription and rearrangements at the T cell
receptor, thereby affecting thymocyte differentiation.\textsuperscript{122}
Cohesins have been shown to mediate long-range transcriptional regulation by controlling the spatial conformation of chromatin at multiple gene loci that are important for normal development and differentiation. Studies revealed two distinct types of cohesin sites: sites that coincide with the binding of CTCF (CCCTC-binding factor) vs. sites that map to active enhancers and promoters and are usually cell-type specific. The CTCF-dependent interaction of cohesins with insulator blocks enhancer activity and disrupts distal enhancer-promoter interactions required for gene activation. Moreover, cohesin has a CTCF-independent role in tissue-specific transcriptional regulation. ChIP-Seq data suggest that cohesins co-localize with master regulators in several tissues, such as liver-specific transcription factors in HepG2 cells and estrogen receptor α in MCF-7 cells. Cohesin also co-localize with transcriptional coactivators, such as mediator to facilitate chromatin looping between the enhancer and promoter of some pluripotency genes (e.g POU5F1) in mouse embryonic stem cells. The cohesin complex lacks a definitive DNA-binding domain. Therefore, DNA localization appears to be facilitated through binding to CTCF and transcription factors, thus forming a regulatory network for transcriptional programs of specific cell type.

Cohesins play an essential role in the maintenance of pluripotency. Depletion of cohesins blocks self-renewal, induces spontaneous differentiation, and interferes with reprogramming of fibroblasts to pluripotent cells. Mutations in core components of the cohesin complex can cause developmental defects in a number of species. For instance, heterozygous mutations in cohesin loader NIPBL or less frequently, in cohesin subunits SMC1A and SMC3 result in Cornelia de Lange syndrome (CdLS), a neurodevelopmental disorder with upper extremity malformations.

1.2.2 Cohesin deregulation in cancer
Mutations of cohesins have been found in many cancers including leukemias\textsuperscript{19, 49, 128}, colorectal carcinomas\textsuperscript{129}, ovarian carcinomas\textsuperscript{11, 130}, glioblastoma, melanomas, and Ewing’s sarcomas\textsuperscript{131}. The first somatic mutations of cohesin in cancer were reported in 2008 when heterozygous missense mutations in \textit{SMC1A}, \textit{SMC3}, \textit{STAG3} (a component of meiotic cohesin) and \textit{NIPBL} were identified in aneuploid colorectal cancers.\textsuperscript{129} In 2010, deletions of \textit{RAD21} in a CML and deletions of \textit{STAG2} in an AML were reported.\textsuperscript{132} In 2011, \textit{STAG2} mutations were reported to result in cohesion defects and aneuploidy in glioblastoma cell lines, melanomas, and Ewing’s sarcomas.\textsuperscript{131} \textit{STAG2} is the most frequently mutated gene of the cohesin complex. Because \textit{STAG2} is located on the X chromosome, only a single mutational event is required to inactivate it in both males and females (due to X inactivation). \textit{STAG2} mutations are considered loss-of-function mutations because: 1) the majority of mutations are truncating, 2) truncating mutations are present in early exons, resulting in a very short protein, 3) in many cases a truncated \textit{STAG2} protein is absent, likely due to nonsense-mediated decay of the mutant \textit{STAG2} mRNA.\textsuperscript{133, 134} In 2013, three studies reported frequent somatic mutations of \textit{STAG2} in bladder cancer.\textsuperscript{135, 136, 137} In addition, \textit{SMC1A} has been shown to be overexpressed in gliomas and reducing its levels inhibits glioma cell growth \textit{in vitro}.\textsuperscript{138} Upregulation of \textit{ESCO2} and \textit{WAPL} is associated with tumor progression in melanomas and cervical cancer, respectively.\textsuperscript{139, 140} Overexpression of Separase is sufficient to induce tumorigenesis in mammary epithelial cells in a \textit{TP53}-mutant background.\textsuperscript{141} Pan-cancer analysis of the TCGA data found that the cohesin complex was recurrently mutated across 12 cancer types and identified the cohesin complex as one of the 16 significantly mutated subnetworks.\textsuperscript{142}

1.2.2.1 Cohesin mutations in myeloid malignancies

\textit{Cohesin mutations in AML}
Recurrent mutations in all four members of the cohesin complex, *SMC1A, SMC3, RAD21*, and *STAG2* were first identified in M1 AML cases. They co-occurred with *NPM1, RUNX1, DNMT3A*, or *TET2* mutations in 17/19 cases, indicating cooperation with other leukemogenic pathways. Subsequently, the TCGA data confirmed and extended these results, identifying a cumulative cohesin mutation frequency of 13% (26/200). The frequency of cohesin mutations was assessed in both *de novo* and secondary AML. One study showed a higher frequency of cohesin mutations in *de novo* AML (13%, 16/120 samples studied) than in secondary AML (8%, 3/37 samples studied). In contrast, another study showed a higher frequency of cohesin mutations in secondary AML (20%, 30/149) than in *de novo* AML (11%, 32/301). In both studies, the most frequently mutated cohesin gene was *STAG2*, followed by *SMC3* and *RAD21*. Cohesin mutations are nearly always mutually exclusive and are mostly found in samples with normal karyotypes. Based on allelic burden analysis, cohesin mutations are often, but not always, observed as early event during leukemogenesis.

*Cohesin mutations in MDS*

*STAG2* mutations were identified in MDS samples from patients whose disease later progressed to secondary AML. Subsequent studies showed that 7% (10/150) of MDS samples harbor *STAG2* mutations and 8% (18/224) of MDS samples harbor cohesin mutations, the majority of which were *STAG2* mutations, with lower mutation frequencies in *SMC3* and *RAD21*. Cohesin mutations were also identified in 17% of high-risk MDS samples and 11% of low-risk samples, respectively.

*Cohesin mutations in DS-AMKL*

Down syndrome is associated with trisomy 21, and individuals with trisomy 21 are more susceptible to hematologic abnormalities. Up to 10% of children with Down syndrome will
present with transient abnormal myelopoiesis at birth, a necessary predecessor to DS-AMKL. Virtually all DS-AMKL patients have an inactivating mutation in \textit{GATA1}, which results in exclusive expression of a shorter isoform, named GATA1s. \textsuperscript{80}

Cohesin mutations are prominent in DS-AMKL and are predicted to be heterozygous, loss-of-function, and early events during leukemia development. Deep sequencing revealed that 53\% of the DS-AMKL samples had acquired cohesin mutations that were not in the self-limiting pre-leukemic transient abnormal myelopoiesis, suggesting cohesin haploinsufficiency may drive oncogenic transformation and progression to DS-AMKL. \textsuperscript{79} In addition to the presence of trisomy 21 and GATA1s, cohesin mutations likely cooperate with chromosome 21 genes such as \textit{RUNX1}, \textit{ERG}, and \textit{ETS2} to promote the development of DS-AMKL. \textsuperscript{80}\textsuperscript{,} \textsuperscript{145} Furthermore, mutations in \textit{CTCF} occur in approximately 20\% of DS-AMKL and are not mutually exclusive to cohesin mutations. \textsuperscript{80} Cohesins and CTCF interact to regulate chromatin architecture, and thus mutations in either could have non-overlapping effects on genomic structure and induce global changes on gene expression.

\textit{Phenotypic consequences of cohesin mutations on hematopoiesis}

Four recent studies sought to elucidate the phenotypic consequences of loss of cohesin and cohesin mutations on hematopoiesis in mouse and human models (Table 1. \textsuperscript{146}\textsuperscript{,} \textsuperscript{147}\textsuperscript{,} \textsuperscript{148}\textsuperscript{,} \textsuperscript{149}\textsuperscript{,} \textsuperscript{150} Viny et al. showed a dose-dependent role for Smc3 in regulating hematopoietic stem and progenitor cell (HSPC) function and chromatin structure. Biallelic loss of \textit{Smc3} in mice led to bone marrow aplasia with premature sister chromatid separation, revealing an absolute requirement for cohesin in HSPC function; whereas, Smc3 haploinsufficiency increased self-renewal \textit{in vitro} and \textit{in vivo}. Furthermore, \textit{Smc3} haploinsufficiency reduced expression of
transcription factors and lineage commitment-associated genes and cooperated with *FLT3-ITD* mutation to induce AML *in vivo*.\textsuperscript{147}

Mullenders et al. generated a series of inducible shRNA mouse models targeting each of the four cohesin subunits. Knockdown of cohesin resulted in gain of replating capacity of mouse HSPCs and altered hematopoiesis with skewing towards myeloid differentiation. Upregulation of genes involved in myeloid differentiation and increased chromatin accessibility around those genes were also observed. In addition, aged cohesin knockdown mice developed a clinical picture closely resembling MPNs, implying that cohesin mutations can occur as an early event in leukemogenesis and facilitate the potential development of a myeloid malignancy.\textsuperscript{148}

Complementary work in cohesin mutant human HSPCs showed that depletion of cohesin subunits increased replating capacity *in vitro* and led to myeloid-skewed differentiation, consistent with phenotypes seen in mouse models. Mazumdar et al. found that introduction of cohesin mutants into AML cell lines and primary human cord blood HSPCs resulted in a differentiation block with an increased frequency of CD34+ cord blood progenitors. Cohesin mutants augmented the serial replating capability of human HSPCs *in vitro* and elevated chromatin accessibility and predicted transcription factor binding for HSPC regulators including RUNX1, GATA2, and ERG, measured by ATAC-Seq and ChIP-Seq.\textsuperscript{149}

Similarly, Galeev et al. identified several members of the cohesin complex SMC3, RAD21, STAG1/2 in an RNAi screen as critical modifiers of self-renewal and differentiation in human HSPCs. They showed that cohesin deficiency induced HSC-specific gene programs and the reconstitution potential of cohesin-deficient HSPCs was increased in primary and secondary transplantation studies.\textsuperscript{150}
Figure Legends

Figure 1.1. Eight functional categories of genes that are frequently mutated in AML.\textsuperscript{17}

Mutations in signaling genes such as $FLT3$ (upper left box). Mutations in tumor-suppressor
genes such as $TP53$ (upper middle box). Mutations in DNA-methylation-associated genes such
as $DNMT3A$, $TET2$, $IDH1$, and $IDH2$ (upper right box). Mutations in myeloid transcription
factors such as $RUNX1$ (center left box). Mutations in cohesin-complex genes such as $STAG2$
and $RAD21$ (center middle box). Mutations in chromatin-modifying genes such as $ASXL1$ and
$PHF6$ (center right box). Mutations in $NPM1$ (lower left box). Mutations in spliceosome-
complex genes such as $SRSF2$, $SF3B1$, $U2AF1$, and $ZRSR2$ (lower right box).

Figure 1.2. The cohesin complex.\textsuperscript{86}

Cohesin is a ring-shaped complex, composed of four core subunits SMC1A, SMC3, RAD21, and
STAG1/2. SMC1A and SMC3 form intramolecular antiparallel coiled coils and fold back on
themselves, creating a hinge domain at one end and an ATPase head at the other. SMC1A and
SMC3 dimerize at the hinge domains and their ATPase heads are bound by RAD21. STAG1/2
interacts with the central region of RAD21. PDS5, SORORIN, and WAPL are regulatory
proteins of cohesin.
Figure 1.1. Eight functional categories of genes that are frequently mutated in AML.\textsuperscript{17}
Figure 1.2. The cohesin complex.\textsuperscript{86}
<table>
<thead>
<tr>
<th>Mammals</th>
<th>D. melanogaster</th>
<th>S. cerevisiae</th>
<th>S. pombe</th>
<th>Function</th>
</tr>
</thead>
<tbody>
<tr>
<td>SMC1A</td>
<td>Smc1</td>
<td>Smc1</td>
<td>Psm1</td>
<td>Core subunit (mitosis)</td>
</tr>
<tr>
<td>SMC1B</td>
<td>Core subunit (meiosis)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>SMC3</td>
<td>Smc3</td>
<td>Smc3</td>
<td>Psm3</td>
<td>Core subunit</td>
</tr>
<tr>
<td>RAD21</td>
<td>Rad21/Vtd</td>
<td>Scc1/Mcd1</td>
<td>Rad21</td>
<td>Core subunit (mitosis)</td>
</tr>
<tr>
<td>REC8</td>
<td>C(2)M</td>
<td>Rec8</td>
<td>Rec8</td>
<td>Core subunit (meiosis)</td>
</tr>
<tr>
<td>STAG1/S1A</td>
<td>SA</td>
<td>Scc3</td>
<td>Psc3</td>
<td>Core subunit (mitosis)</td>
</tr>
<tr>
<td>STAG2/S2A</td>
<td>SA2</td>
<td>Scc3</td>
<td></td>
<td>Core subunit (mitosis)</td>
</tr>
<tr>
<td>STAG3/S3A</td>
<td>/</td>
<td>Rec11</td>
<td></td>
<td>Core subunit (meiosis)</td>
</tr>
<tr>
<td>NIPBL/SCC2</td>
<td>Nipped-B</td>
<td>Scc2</td>
<td>Mis4</td>
<td>Cohesin loading</td>
</tr>
<tr>
<td>MAU2/SCC4</td>
<td>Scc4</td>
<td>Scc4</td>
<td>Ssl3</td>
<td>Cohesin loading</td>
</tr>
<tr>
<td>ESCO1</td>
<td>Eco/Deco</td>
<td>Eco1/Ctf7</td>
<td>Eso1</td>
<td>Cohesion establishment</td>
</tr>
<tr>
<td>ESCO2</td>
<td>San</td>
<td>Eco1/Ctf7</td>
<td></td>
<td>Cohesion establishment</td>
</tr>
<tr>
<td>PDS5A</td>
<td>Pds5</td>
<td>Pds5</td>
<td>Pds5</td>
<td>Cohesion maintenance</td>
</tr>
<tr>
<td>PDS5B</td>
<td></td>
<td></td>
<td></td>
<td>Cohesion maintenance</td>
</tr>
<tr>
<td>WAPL</td>
<td>Wap1</td>
<td>Wpl1/Rad61</td>
<td>Wpl1</td>
<td>Cohesion maintenance</td>
</tr>
<tr>
<td>SORORIN/CDC5</td>
<td>Dalmatian</td>
<td>/</td>
<td></td>
<td>Cohesion maintenance</td>
</tr>
<tr>
<td>HDAC8</td>
<td>/</td>
<td>Hos1</td>
<td>/</td>
<td>Cohesin dacetylase</td>
</tr>
<tr>
<td>Shugosin1/SGOL1</td>
<td>Mei-S332</td>
<td>Sgo1</td>
<td>Sgo1</td>
<td>Cohesin protection</td>
</tr>
<tr>
<td>Separase</td>
<td>Sse1</td>
<td>Esp1</td>
<td>Separase</td>
<td>Cohesin removal</td>
</tr>
<tr>
<td>Polo like Kinase 1 (PLK1)</td>
<td>Polo</td>
<td>Cdc5</td>
<td>Plk1</td>
<td>Cohesin removal</td>
</tr>
</tbody>
</table>
Table 1.2. Studies of cohesin mutations in hematopoiesis.\textsuperscript{146}

<table>
<thead>
<tr>
<th>Model system</th>
<th>Approach</th>
<th>Conclusions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mouse model</td>
<td>Smc3 biallelic and haploinsufficient conditional knockout</td>
<td>Increased replating, enrichment of HSPC gene signature, chromatin accessibility changes, dose dependent</td>
</tr>
<tr>
<td>Mouse model</td>
<td>shRNA knockdown of cohesin subunits</td>
<td>Increased replating, enrichment of HSPC gene signature, chromatin accessibility changes, MPN-like phenotype in aged mice</td>
</tr>
<tr>
<td>Human cord blood (HSPCs)</td>
<td>Lentiviral transduction of cohesin mutants or shRNA knockdown</td>
<td>Increased replating, enrichment of HSPC gene signature, chromatin accessibility changes</td>
</tr>
<tr>
<td>Human cord blood (HSPCs)</td>
<td>RNAi screen</td>
<td>Increased replating, enrichment of HSPC gene signature, increased secondary transplant engraftment</td>
</tr>
</tbody>
</table>
References


Chapter 2:

*Smc3* is required for mouse embryonic and adult hematopoiesis
Abstract

SMC3 encodes a subunit of the cohesin complex that has canonical roles in regulating sister chromatids segregation during mitosis and meiosis. Recurrent heterozygous mutations in SMC3 have been reported in acute myeloid leukemia (AML) and other myeloid malignancies. In this study, we investigated whether the missense mutations in SMC3 might have dominant-negative effects or phenocopy loss-of-function effects by comparing the consequences of Smc3 deficient and haploinsufficient mouse models. We found that homozygous deletion of Smc3 during embryogenesis or in adult mice led to hematopoietic failure, suggesting that SMC3 missense mutations are unlikely to be associated with simple dominant negative phenotypes. In contrast, Smc3 haploinsufficiency was tolerated during embryonic and adult hematopoiesis. Under steady-state conditions, Smc3 haploinsufficiency did not alter colony forming in methylcellulose, only modestly decreased mature myeloid cell populations, and led to limited expression changes and chromatin alteration in Lin-cKit+ bone marrow cells. However, following transplantation, engraftment, and subsequent deletion, we observed a hematopoietic competitive disadvantage across myeloid and lymphoid lineages and within the stem/progenitor compartments. This disadvantage was not affected by hematopoietic stresses but was partially abrogated by concurrent Dnmt3a haploinsufficiency, suggesting that antecedent mutations may be required to optimize the leukemogenic potential of Smc3 mutations.

Introduction

Acute myeloid leukemia (AML) is an aggressive hematopoietic malignancy, characterized by the accumulation of myeloblasts in the blood or bone marrow (BM) with maturation arrest and retained self-renewal. Tremendous progress has been made in identifying recurrent gene
mutations in AML, yet we are still in the early stages of understanding the mechanisms through which these genetic alterations contribute to the onset of the disease.\(^2\)

Recurring mutations in the cohesin complex occur in four core components, \textit{SMC3}, \textit{SMC1A}, \textit{RAD21}, and \textit{STAG2}, and have been identified in AML and other myeloid malignancies.\(^3,4,5\) Over 50\% of patients with Down syndrome-associated acute megakaryocytic leukemia (DS-AMKL) have cohesin mutations, specifically in \textit{STAG2}.\(^6\) Somatic cohesin mutations have also been observed in a variety of solid cancers, including colorectal carcinoma, ovarian carcinoma, glioblastoma, bladder carcinoma, and Ewing’s sarcoma.\(^7,8,9,10,11,12\) Additionally, germline mutations of the cohesin complex are causally related to developmental disorders, particularly cohesinopathies such as Cornelia de Lange syndrome (CdLS).\(^13,14\)

\textit{SMC3} and \textit{RAD21} mutations are nearly universally heterozygous, whereas mutations in \textit{SMC1A} and \textit{STAG2} may be hemizygous because they are X-linked. Cohesin mutations also tend to be mutually exclusive, implying that alteration in one component may be sufficient to disrupt the entire complex or alternatively, they may not be tolerated by a cell when co-occurring.\(^15,16\)

Cohesin mutations are often observed as early subclonal events in AML, conceivably facilitating disease initiation, although they are not observed in cases of clonal hematopoiesis of indeterminate potential (CHIP), suggesting they are unlikely to be the initiating event.\(^15,17,18,19\)

The majority of \textit{SMC3} mutations are missense mutations; only one-third of \textit{SMC3} mutations are nonsense or splice-site variants. The missense mutations are scattered across all domains, although a few recurrently mutated nucleotides have been observed (R381Q, R661P). This pattern suggests that many of these mutations may result in simple loss-of-function consequences, although novel dominant negative activities cannot be dismissed within the hot-spot variants. Intriguingly, \textit{DNMT3A} mutations, one of the most commonly mutated genes in
AML, frequently coincided with *SMC3* mutations, suggesting there may be leukemogenic interactions between these mutations.\(^5, 15, 16, 20\)

In yeast and cell line-based studies, cohesin has been shown to play essential roles in sister chromatid segregation during cell cycle, DNA damage repair, transcriptional regulation via chromatin looping, and maintenance of chromatin architecture.\(^21, 22, 23, 24\) Notably, AML patients who harbor cohesin mutations typically have normal karyotype, indicating that hematopoietic cohesin mutations do not lead directly to chromosomal instability.\(^25, 16\)

To define the hematopoietic consequences of *SMC3* mutations and to determine whether these could reflect dominant negative or loss of function phenotypes, we characterized the *in vivo* effects of *Smc3* deficiency and *Smc3* haploinsufficiency on murine hematopoiesis using conditionally deleted strategies. In contrast to our expectations that these leukemia-associated mutations would lead to expansions of hematopoietic stem cell populations or augmented self-renewal, we observed a competitive disadvantage in *Smc3* deficient and haploinsufficient BM cells *in vivo* without an associated increase in maturation-arrested stem cells.

**Methods**

**Animal Studies**

*Smc3*\(^{trap}\) mice were obtained from the European Conditional Mouse Mutagenesis Program (EUCOMM) (Smc3<tm1a(EUCOMM)Wtsi>, MGI:4434007). To generate *Smc3*\(^{fl}\) mice, the gene-trap was removed by crossing *Smc3*\(^{trap}\) mice with Flp deleter mice (B6.129S4-Gt(Rosa)26Sortm2(FLP*)Sor/J), and subsequently outbreeding the Flp allele with C57BL/6J intercrosses. We generated *Smc3* conditional deficient mice by breeding the *Smc3*\(^{fl/fl}\) mice with *Vav1-Cre* (B6.Cg-Commd10Tg(Vav1-cre)A2Kio/J), *ERT2-Cre* (B6.Cg-Tg(cre/Esr1)5Amc/J),
and CMV-Cre (B6.C-Tg(CMV-cre)1Cgn/J), obtained from the Jackson Laboratory. We characterized Smc3 conditional deficient mice at 6-8 weeks old and both genders were used. Whenever possible, littermate controls were used for all experiments. CBCs were measured using Hemavet 950 (Drew Scientific Group).

All mice were on the C57BL/6 background and were cared for in the Experimental Animal Center of Washington University School of Medicine. The Washington University Animal Studies Committee approved all animal experiments.

**Intracellular Smc3 staining**

Intracellular Smc3 was detected with the Pharmingen™ Transcription Factor Buffer Set (562574 BD Biosciences) according to the manufacturer’s instructions. BM cells were isolated from femurs and tibias and lysed with ACK lysis buffer (150mM NH₄Cl, 10mM KHCO₃, 0.1mM Na₂EDTA [Na₂-ethylenediaminetetraacetic acid], PH7.2-7.4). Cells were stained with cell-surface markers to identify cell type by flow cytometry and then fixed for 40 minutes at 4°C. Cells were washed with perm wash buffer and incubated with primary antibody against Smc3 (1:100 dilution, ab9263, Abcam) for 30 minutes at 4°C. Cells were washed in perm wash buffer and incubated in secondary antibody (1:500 dilution, chicken anti-rabbit Alexa Fluor 647, Molecular Probes) for 30 minutes at 4°C. Cells were rinsed in perm wash buffer and analyzed by flow cytometry. The mean fluorescence intensity was calculated for the AF647 signal.

**Flow cytometry**

After lysis of red blood cells by ACK lysis buffer, peripheral blood, BM, spleen cells, or thymocytes were treated with anti-mouse CD16/32 (eBioscience; clone 93) and stained with the indicated combinations of the following antibodies (all antibodies are from eBioscience unless
noted otherwise): CD34 FITC (clone RAM34), CD11b PE (clone M1/70), c-Kit PECy7 (clone 2B8) or BV421 (BioLegend, clone 2B8), Sca1 PE-Dazzle™ 594 (BioLegend, clone D7) or APC (clone D7), Gr-1 FITC, PECy7, APC (clone RB6-8C5), or BV421 (BioLegend, clone RB6-8C5), B220 PE, PECy7, APC (clone RA3-6B2), or APC-Cy7 (BioLegend, clone RA3-6B2), CD3 PECy7 (clone 145-2C11), CD71 PE (clone R17217), Ter-119 PECy7 or APC (clone TER-119), CD16/32 BV510 (clone 93), CD150 PE (BioLegend 115903, clone TC15-12F12.2), CD48 APC-Cy7 (BioLegend, clone HM48-1), Ly5.1 APC (clone A20) or AF700 (BioLegend, clone A20), Ly 5.2 PE or e450 (clone 104). The following flow phenotypes were used for stem and progenitor cell flow: Lin- (lineage negative): B220-, CD3e-, Gr-1-, Ter-119-, CD4-, CD8-, CD19-, CD127-; KL: Lin-, cKit+, Sca1-; KLS: Lin-, cKit+, Sca-1+; KLS-SLAM: Lin-, cKit+, Sca-1+, CD150+, CD48--; GMP: Lin-, cKit+, Sca-1-, CD34+, CD16/32+; CMP: Lin-, cKit+, Sca-1-, CD34+, CD16/32--; and MEP: Lin-, cKit+, Sca-1-, CD34-, CD16/32-.

Analysis was performed using a FACScan (Beckman Coulter) or Gallios flow cytometer (Beckman Coulter). Cell sorting was performed using I-Cyt Synergy II sorter (I-Cyt Technologies). Flow cytometry data were analyzed with FlowJo Software Version 10 (TreeStar), Excel (Microsoft), and Prism 7.02 (GraphPad Software).

**Competitive transplantation**

Competitive transplantation was performed using 0.5 x 10^6 whole BM cells from indicated donor mice (CD45.2) mixed with 0.5 x 10^6 competitor whole BM cells wild-type CD45.1 (Ly5.1) x CD45.2 mice. Mixture cells were injected intravenously into 6-8 weeks old CD45.1 recipient mice that received 1,100 cGy total body irradiation (Mark 1 Cesum irradiator, J.L. Shepard) 24 hours prior to transplantation. For Smc3^fl/fl/ERT2-Cre^+/− or Smc3^fl/+/ERT2-Cre^+/− transplantation, recipient mice were treated with tamoxifen (dissolved in sterile corn oil, Sigma-Aldrich) 6 weeks
post-transplant via oral gavage for 9 doses (3 mg/day/mouse, 3 days/week). Peripheral blood was examined for donor cell chimerism at indicated time points after transplantation. Recipient mice BM were analyzed at the end of experiment.

**Colony replating assay**

BM cells were harvested and plated in duplicate (10,000 BM cells/plate) in complete mouse methylcellulose medium with stem cell factor, IL-3, IL-6, and Epo (R&D Systems). Colonies were counted on day 7, and cells were collected from methylcellulose in warm Dulbecco modified Eagle medium containing 2% fetal bovine serum, washed, and replated as before. An aliquot of cells was taken for analysis of myeloid (Gr1, CD11b) and mast cell markers (cKit, FcER1) by flow cytometry. This process was repeated for 4 weeks or until colony formation failed.

**RNA sequencing of multipotent progenitors and analysis**

Multipotent progenitors (KLs; Lin-, cKit+, Sca1-) were sorted from three wild-type or Smc3fl/+Vav1-Cre+/- mice into DMEM media. Flow cytometry of samples after sorting validated >93% sort accuracy. RNA was extracted from cell pellets using a miRNeasy kit (QIAGEN) and genomic DNA was removed by RNase-Free DNase Set (QIAGEN). RNA was analyzed for degradation using the RNA Nano Chip (Agilent #5067-1521). An input of 300ng was taken forward for each sample using the TruSeq Stranded Total RNA with Ribo-Zero Globin Kit (Illumina #20020612). Final Libraries were analyzed using the High Sensitivity DNA Chip (Agilent# 5067-4626). All Libraries were pooled and run across 3 lanes of HiSeq4000. RNAseq data were aligned to the human reference with Tophat v2.0.8 (denovo mode, params: --
library-type fr-firststrand --bowtie-version=2.1.0). Expression levels were calculated with Cufflinks v2.1.1 (params: --max-bundle-length 10000000 --max-bundle-frags 10000000). 52

**ATAC- Sequencing of multipotent progenitor and analysis**

Chromatin accessibility assays using the bacterial Tn5 transposase were performed using multipotent progenitors (KLs; Lin-, cKit+, Sca1-) sorted from Smc3^{3/2} or Smc3^{3/2}/Vav1-Cre^{+/−} mice in triplicate. DNA was prepared from 75,000 sorted cells and >93% sorting accuracy verified with post-sort analysis. ATAC libraries were generated exactly as described 53 and pooled and sequenced on a HiSeqX instrument (Illumina) to obtain between 133 and 152 million 2x150 bp paired-end reads. Raw sequencing reads were adapter trimmed with trim galore using cutadapt version 1.8.1 (Martin EMBnet 2011) and then aligned to the mouse reference genome (mm10) using bwa mem (Li H. arXiv:1303.3997v1 (2013)). Peaks in each sample were identified with macs2 54 using the -f BAMPE parameter and then filtered to retain peaks with a q-value <0.01. Peak summits from all samples were merged together with BEDtools merge 55 using parameters to combine summits within 50 bp of each other. Read counts at the merged peak summits were obtained for all samples using the deepTools multiBamSummary command 56 with the minimum mapping quality set to 1, and then processed using DESeq2 57 with default parameters to obtain normalized counts for each peak summit and to perform differential analysis across all peaks between wild-type and mutant mice.

**Statistics**

Statistical analysis was performed using Prism 7.02 (GraphPad Software) and Excel (Microsoft). Unpaired two-tailed t-test, one-way, and two-way ANOVA with Turkey’s multiple comparisons tests were performed, as appropriate. P values < 0.05 were considered statistically significant.
Error bars represent standard deviation. Data points without error bars have standard deviation below Prism 7.02’s limit to display.

Results

Generation of Smc3 conditional knockout mice

To investigate the effects of Smc3 loss on hematopoiesis, we generated Smc3 conditionally deficient and haploinsufficient mice using Smc3<tm1a(EUCOMM)Wtsi> mice obtained from EUCOMM (Smc3<trap>). The Smc3<trap> allele has a lacZ-neomycin-gene-trap cassette inserted in intron 4 with two Frt sites on each side of the cassette, and two loxP sites flanking exon 4. The gene trap is predicted to lead to an early transcription stop after splicing into lacZ-neomycin. The conditional knockout Smc3<fl> allele was created by excising the gene-trap cassette with Flp recombinase and was used for further characterizations because homozygous deletion could be achieved using the Smc3<fl> allele (Figure 2.1A). We validated the integration of the loxP sites surrounding exon 4 in the Smc3<fl> allele using whole genome sequencing (Figure 2.1B).

We examined the transcriptional consequences of the Smc3<fl> allele using RNA-Seq and intracellular flow cytometry. In BM cells from three Smc3<fl>/Vav1-Cre<+/> mice, nearly 50% (48.4%) of transcripts spliced from exon 3 to exon 5, consistent with deletion of exon 4 while all the wild-type transcripts spliced from exon 3 to exon 4 and exon 4 to exon 5 (Figure 2.1C and Figure 2.7A-F). Analysis of reads spanning exons 3-5 suggests that this results in a frameshift mutation and a stop codon after 59 amino acids, although this truncated protein could not be detected using N-terminal antibodies. Using C-terminal antibodies, intracellular Smc3 protein level was reduced to approximately half of littermate control, as would be expected with a heterozygous allele and confirming Smc3 haploinsufficiency (Figure 2.1D). In addition, we noted
that Smc3 protein level was regulated during normal hematopoiesis, with higher expression in KLS (Lin-cKit+Sca1+) stem/progenitor cells vs. SLAM (Lin-cKit+Sca1+CD150+CD48-) stem cells (Figure 2.1E). Representative primary intracellular flow data shown in Figure 2.8.

**Homozygous Smc3 deletion**

To understand whether SMC3 mutations might have dominant-negative effects or phenocopy loss-of-function effects, we compared the consequences of Smc3 deficient and haploinsufficient mouse models. We found that hematopoietic homozygous deletion of Smc3 led to embryonic lethality. In heterozygous Smc3<sup>fl/fl</sup>/Vav1-Cre<sup>+/−</sup> intercrosses, we observed 0 out of 75 pups with homozygous Smc3 alleles (Figure 2.2A). To determine whether the cause of death in Smc3<sup>fl/fl</sup>/Vav1-Cre<sup>+/−</sup> embryos was from hematopoietic failure, we examined E13.5 embryos. Grossly, the Smc3<sup>fl/fl</sup>/Vav1-Cre<sup>+/−</sup> embryos were indistinguishable in size and appearance from other genotypes, except the lack of obvious fetal livers (Figure 2.2B-C). A severe decrease in fetal liver hematopoietic cells was verified by cell count and flow cytometry with near-complete absence of CD45+ Gr1+ CD11b+ cells demonstrating myeloid-biased hematopoietic failure (Figure 2.2D-F).

We investigated somatic homozygous Smc3 deletion in adult mice using the Smc3<sup>fl/fl</sup>/ERT2-Cre<sup>+/−</sup> mice. Smc3 deletion was achieved by treating mice with oral tamoxifen (TAM) at 6 weeks of age and reduction in Smc3 protein confirmed with western blot (Figure 2.9A). After 4 doses of TAM, mice were moribund and therefore sacrificed for analysis. Complete blood counts (CBC) data showed the Smc3<sup>fl/fl</sup>/ERT2-Cre<sup>+/−</sup> mice had lower white blood cell counts, percentages of lymphocytes and monocytes, and fewer platelets than TAM-treated littermates (Figure 2.3A). The Smc3<sup>fl/fl</sup>/ERT2-Cre<sup>+/−</sup> mice had decreased spleen weights (Figure 2.3B) and their spleens were smaller in size (Figure 2.9B). Total number of cells in BM, spleen, and
thymus of the $\text{Smc3}^{0/0}/\text{ERT2-Cre}^{+/−}$ mice were significantly reduced in comparison to $\text{Smc3}^{0/0}$ mice after TAM treatment (Figure 2.3C). The reduction of cells occurred across all lineages in the BM (Figure 2.3D), spleen, and thymus (Figure 2.9C-D) of the $\text{Smc3}^{0/0}/\text{ERT2-Cre}^{+/−}$ mice, suggesting complete hematopoietic collapse.

Because activation of ERT2-Cre leads to $\text{Smc3}$ deletion in a wide range of cells and tissues, we repeated these studies, isolating hematopoietic cells via a competitive transplantation. Equivalent engraftment of transgenic CD45.2+ and competitor CD45.1+ CD45.2+ cells was verified 6 weeks after transplantation. Following tamoxifen-induced $\text{Smc3}$ deletion, the $\text{Smc3}^{0/0}/\text{ERT2-Cre}^{+/−}$ donor cells were quickly outcompeted, indicating complete loss of hematopoietic stem and progenitor cell (HSPC) functions in the $\text{Smc3}^{0/0}/\text{ERT2-Cre}^{+/−}$ BM. Once again, the effect was most pronounced within the myeloid compartment (Figure 2.3E-F), suggesting that myeloid hematopoiesis is sensitive to $\text{Smc3}$ deletion and therefore, the AML-associated $\text{SMC3}$ mutations are unlikely to have simple dominant-negative effects.

**Steady-state heterozygous $\text{Smc3}$ deletion**

In the ExAC database (exac.broadinstitute.org), no $\text{SMC3}$ loss-of-function mutations are observed in available human data (0 observed vs. 58.5 expected mutations), suggesting potential embryonic lethality or reduced fitness associated with $\text{Smc3}$ haploinsufficiency. We, therefore, determined whether $\text{Smc3}$ haploinsufficiency might be tolerated in mice. Because $\text{CMV-Cre}$ is X-linked and expressed during early embryogenesis, we examined the ratio of male: female pups and compared difference between genders to determine whether embryonic $\text{Smc3}$ haploinsufficiency altered hematopoiesis. We found that $\text{Smc3}$ haploinsufficiency led to a normal number of female pups in $\text{CMV-Cre}$ intercrosses (Figure 2.10A), and the female pups had no obvious defects in complete blood counts, total number of BM cells, and percentages of HSPCs
and cells in different lineages (Figure 2.10B-E). Hence, embryonic $Smc3$ haploinsufficiency could be tolerated and did not grossly perturb steady-state hematopoiesis in mice.

We next assessed the effects of somatic $Smc3$ haploinsufficiency on hematopoiesis using the inducible $Smc3^{0/+}/ERT2-\text{Cre}^{+/−}$ mice. $Smc3$ haploinsufficiency did not alter the proportions of immunophenotypic HSPCs and cells of different lineages (Figure 2.4A-B).

Furthermore, $Smc3$ haploinsufficiency did not increase the number of colonies formed in methylcellulose or the average number of cells per colony, and the $Smc3$ haploinsufficient BM cells did not replate beyond two weeks (Figure 2.4C-E). At the end of each week, the colonies on each plate were collected, washed, and characterized by immunophenotype. At the end of week 1, the cells were predominantly $\text{Gr}1^{+}\text{CD}11b^{+}$ for both $Smc3^{0/+}/ERT2-\text{Cre}^{+/−}$ and $Smc3^{0/+}$ genotypes. However, starting week 2, the colonies shifted to $\text{cKit}^{+}\text{FcER1}^{+}$ mast cells. In week 3 and 4, the few colonies left were exclusively mast cells (Figure 2.11A-B). Similar results were observed using BM cells from $Smc3^{0/+}/\text{Vav1-Cre}^{+/−}$ mice.

We performed RNA-Sequencing to measure global gene expression in $Smc3$ haploinsufficient hematopoietic progenitors ($\text{Lin-Kit}^{+}\text{Sca1}^{−}$) using the constitutive $Smc3^{0/+}/\text{Vav1-Cre}^{+/−}$ model. This model was chosen because it required minimal manipulation of the mice, provided hematopoietic-restricted deletion, and would evaluate steady-state hematopoietic conditions. Multipotent progenitors (KLS) were sorted from age-matched individual wild-type and $Smc3^{0/+}/\text{Vav1-Cre}^{+/−}$ mice for RNA-Seq. KLS were selected because $Smc3$ haploinsufficiency resulted in severe multi-lineage competitive disadvantage in vivo, suggesting potential defect in the functions of $Smc3$ haploinsufficient KLS. However, minimal global transcriptional changes were detected. Using t-tests and Significance Analysis of Microarrays (SAM) $^{26}$ 149 genes were identified with differential expression in $Smc3$ haploinsufficient KLS in comparison to wild-type
controls (most with < 2 fold changes) (Figure 2.4F). KEGG pathway analysis showed significance (p<0.002 and p<0.005) for progesterone mediate oocyte maturation and toxoplasmosis respectively, but these are not related to hematopoiesis.27 Smc3 expression was not observed to be different when analyzed using total reads across the entire gene. However, we observed a two-fold reduction in expression of exon 4 consistent with deletion of this exon (Figure 2.1C).

To determine whether Smc3 haploinsufficiency might lead to alterations in global chromatin structure that may be biologically relevant, but which did not lead to measurable altered gene transcription, we performed transposase-accessible chromatin sequencing (ATAC-Seq). Chromatin accessibility peaks of the Smc3fl/+Vav1-Cre+/KLs and littermate Smc3fl/+controls revealed by ATAC-Seq were not significantly different except for peaks in proximity of three genes: Vav1, Tnpo3, Tgfb (Figure 2.4G). The two-fold difference in Vav1 was expected for the heterozygous allele and therefore indicated fidelity of data generated by the assay.

**Phenotypes of Smc3 haploinsufficiency following competitive transplantations**

AML emerges following clonal expansion. Thus, we conducted competitive transplantation using the inducible ERT2-Cre model instead of the constituent hematopoietic Vav1-Cre, so that complete engraftment could be verified 6 weeks after transplant prior to deletion of the Smc3 allele. In competitive transplantations, we observed a significant competitive disadvantage in the Smc3fl/+ERT2-Cre+/- BM cells (Figure 2.5A). Endpoint analysis of BM cells also showed competitive disadvantage in the Smc3fl/+ERT2-Cre+/- HSPCs and across B and T-cell lineages, implying impaired HSPC functions due to Smc3 haploinsufficiency in the BM, and not a defect in hematopoietic peripheralization or maturation (Figure 2.5B-F). The competitive disadvantage was observed first in the Gr1 myeloid compartment, perhaps due to higher turn-over of these
cells (Figure 2.5D). To verify the competitive disadvantage observed was not due to toxicity of ERT2-Cre, we repeated the competitive transplantation with the ERT2-Cre+/− control mice. The chimerisms of overall CD45.2+ cells and of CD45.2+ cells in all lineages were well-preserved, eliminating the possibility of ERT2-Cre toxicity (Figure 2.12A-B).

The absence of pre-leukemic delayed maturation or augmented self-renewal in Smc3 haploinsufficient mice was unexpected. Hence, we determined whether Smc3 haploinsufficiency might increase self-renewal if it occurred in combination with specific conditions of hematopoietic stress. We again observed a competitive disadvantage in the Smc3fl/+ERT2-Cre+/− BM cells following tamoxifen induction. Intriguingly, the significant myeloid competitive disadvantage was ameliorated at 18 weeks post-transplantation in the pIpC-treated group, whereas it was accelerated in the 5-fluorouracil (5-FU) treated group, although this effect was transient and by week 26 the donor cell population were equivalently reduced (Figure 2.5G-J).

**Dnmt3a haploinsufficiency partially abrogated myeloid competitive disadvantage in Smc3 haploinsufficient BM cells**

In AML patients, DNMT3A mutations co-occurred in approximately one-third of the cases with SMC3 mutations that assess additional mutations.4, 5, 15, 16 We therefore asked whether Smc3 haploinsufficiency might lead to a competitive advantage if it occurred in the background of Dnmt3a haploinsufficiency.20

We observed that with the addition of Dnmt3a haploinsufficiency, the severe competitive disadvantage was partially abrogated in the Smc3fl/+ERT2-Cre+/− myeloid cells, but the significant competitive disadvantage in other lineages remained intact (Figure 2.6A-D). The same phenotype was observed in the Smc3fl/+ERT2-Cre+/−Dnmt3a+/− BM upon endpoint analysis.
 Accordingly, even with constitutive \textit{Dnmt3a} haploinsufficiency, \textit{Smc3} haploinsufficiency did not result in competitive growth advantage in hematopoietic cells.

\textbf{Discussion}

AML is a genetically heterogenous disease characterized by clonal expansion of immature myeloblasts associated with recurrent mutations including the cohesin complex.\textsuperscript{25, 4, 5, 15, 16, 28} Mutations in the subunits of the cohesin complex, \textit{SMC1A}, \textit{SMC3}, \textit{RAD21}, and \textit{STAG2}, have been found as early subclonal events in AML, although they are not observed in people with CHIP.\textsuperscript{5, 15, 16, 18, 19} In contrast, \textit{DNMT3A} mutations are among the most common initiating mutations in normal karyotype AML patients and the most frequently mutated genes in subjects with CHIP.\textsuperscript{25, 29} Cohesin mutations are mutually exclusive of one another and fall into two general categories: mutations in \textit{RAD21} and \textit{STAG2} are mainly truncations and frameshifts, whereas the majority of mutations in \textit{SMC1A} and \textit{SMC3} are missense. In AML, cohesin mutations are not associated with genomic instability, complex karyotypes, or monosomy karyotypes, suggesting alternative pathologic mechanisms.\textsuperscript{4, 5, 15}

To understand whether leukemia-associated \textit{SMC3} missense mutations might have dominant-negative activities or phenocopy loss-of-function effects, we compared the consequences of \textit{Smc3} deficiency and \textit{Smc3} haploinsufficiency on murine hematopoiesis using conditionally deleted strategies. We began by validating the \textit{Smc3} allele using whole genome sequencing, RNA-Seq, and intracellular flow cytometry, which demonstrated correct integration, splicing of approximately 50\% of alleles around exon 4 leading to a frameshift mutation and an early nonsense mutation, and reduced protein levels. Our findings suggest that leukemia-associated \textit{SMC3} mutations are unlikely to have novel dominant negative activities because homozygous \textit{Smc3} deletion was incompatible with embryonic (Figure 2.2) or adult hematopoiesis (Figure
2.3). In these experiments, we observed the effects first in the myeloid compartment. However, because myeloid cells have a shorter half-life than other hematopoietic cell types, the augmented temporal phenotypes observed in these cell fractions may be influenced by greater turn-over. Collectively, these studies demonstrate that *Smc3* is indispensable for embryonic and adult hematopoiesis and normal HSPC functions. Similar severe consequences for *Smc3* deficiency\textsuperscript{30} and *Rad21* deficiency\textsuperscript{31} have been observed, and thus cohesin genes appear to be essential in hematopoietic cells.

Leukemia-associated *SMC3* mutations are observed across all domains of the protein, and nearly one third are nonsense or splice-site variants, suggesting that many of these mutations are likely to be associated with loss of function. Therefore, we investigated the effects of *Smc3* haploinsufficiency on murine hematopoiesis. Because these mutations are associated with leukemia, we predicted that *Smc3* haploinsufficiency would augment colony forming capacity and provide hematopoietic cells a competitive advantage. However, we observed neither phenotype. Following *Smc3* haploinsufficiency induced with three different Cre models (*CMV-Cre*, *Vav1-Cre*, and *ERT2-Cre*) we observed normal CBCs, normal bone marrow hematopoietic population distributions, and normal colony forming (Figure 2.4A-E). We further examined expression signatures and ATAC-Seq under these steady-state conditions in *Vav1-Cre* mice where hematopoietic cells have consistently undergone heterozygous deletion and external perturbations are minimized; we observed little global dysregulation of gene expression or chromatin structure (Figure 2.4F-G). In both studies, internal markers (*Smc3* expression and peaks within the *Vav1* locus) served as controls and markers of the expected dynamic range.

In contrast, under conditions of chimeric competition, *Smc3* haploinsufficiency actually led to competitive disadvantage *in vivo*, with progressive population loss over time (Figure 2.5A-F). In
these studies, Smc3 deletion was induced using ERT2-Cre following a period of 6 weeks post-transplant to facilitate engraftment and stem cell homeostasis prior to deletion. Under these conditions, activation of ERT2-Cre alone does not lead to stem cell toxicity and competitive disadvantage (Figure 2.12A-B), whereas activation of ERT2-Cre just prior to transplantation does.\textsuperscript{32} Analysis of BM populations at the end of the study suggested reduction of populations with Smc3 haploinsufficiency across progenitor and mature cell types, eliminating the possibility that Smc3 haploinsufficiency led to a profound maturation block that prevented leukocyte peripheralization. The competitive disadvantage induced by somatic Smc3 acquisition was unexpected. Therefore, we determined whether specific forms of hematopoietic stress might enable a competitive advantage that could facilitate stem cell expansion and ultimately enable leukemogenesis. We again observed a competitive disadvantage that persisted following a stem cell stressor (5-FU exposure) and an inflammatory stressor (pIpC exposure) (Figure 2.5G-J).

Finally, because SMC3 mutations may not be the first acquired mutation during leukemogenic chronicity, we investigated whether Dnmt3a haploinsufficiency might facilitate Smc3 phenotypes. Germline Dnmt3a haploinsufficiency partially abrogated the myeloid competitive disadvantage of somatically acquired Smc3 haploinsufficiency (Figure 2.6), suggesting that SMC3 mutations may require pre-existing cooperating mutations to facilitate their action. Additionally, these studies do not eliminate the possibility that the frequently observed SMC3 missense mutations may possess novel gain-of-function activity not accessed in these Smc3 haploinsufficient studies.

Thus, under conditions of homeostasis, where all hematopoietic cells have Smc3 haploinsufficiency, murine Smc3 haploinsufficiency does not appear to grossly dysregulate hematopoietic feedback mechanisms or alter normal hematopoietic maturation or self-renewal.
*ex vivo.* However, under conditions of competitive transplantation, we observed a disadvantage in hematopoietic cells across both myeloid and lymphoid lineages suggesting reduced cell production at a multipotent progenitor level.

These results contrast with previously published work using either knock-down strategies in CD34+ cord blood cells or using Mx1-Cre activation with pIpC. Specifically, knocking down of Smc3 using shRNA or RAD21 and SMC1A mutants have been shown to increase self-renewal in human cord blood CD34+ HSPCs *ex vivo*.\(^{33,34}\) Smc3 haploinsufficiency induced by Mx1-Cre exhibited shifts in hematopoietic cell populations, colony forming, and competitive transplantation advantage when deleted using Mx1-Cre two weeks after transplantation\(^ {30}\). These data suggest that differences in the models may interact with the biological consequences of Smc3 reduction through yet undefined mechanisms.

In addition, it is worth noting that other MDS or AML-associated mutations such as U2AF1\(^ {35,36}\), SRSF2\(^ {37,38}\), SF3B1\(^ {39,40,41,42,43}\), ASXL1\(^ {44,45}\) are associated with having competitive disadvantage, which may seem counterintuitive for recurring leukemia mutations observed in patients, but appears to be recurrent biology.

The observed defects in hematopoietic cells with Smc3 deficiency and haploinsufficiency may reflect population data from the ExAC database, where germline cohesin mutations are observed at lower than expected frequencies, suggesting a significant disadvantage in population fitness. No loss-of-function variants are detected in SMC3, SMC1A, STAG2, or RAD21 (based on statistical models of case numbers and gene size, the expected numbers of loss-of-function variants were 58.5, 32, 42.7, and 21.8, respectively). Missense variants were also significantly underrepresented in SMC3, SMC1A, and STAG2, but not in RAD21 (\(z = 6.25, 6.59, 5.11,\) and 2.76; more positive scores indicate fewer variants observed than expected). Of the published
AML-associated missense mutations, only 1 is reported in ExAC (K795E occurring in 3/121,384 alleles), although synonymous changes (R155R, Q367Q, R391R), and alternative amino acid changes (N604S and I1001L) are noted.46

In recent decades, mutations in cohesin complex genes have been associated with genetic syndromes, referred to as cohesinopathies. Several important features differ between cohesinopathies and AML-associated cohesin mutations. Mutations associated with cohesinopathy tend to be in cohesin adapter proteins, such as NILS, HDAC8, and ESCO2, with fewer mutations observed in SMC3, SMC1A, RAD21, or STAG2.47 Cohesinopathies are associated with facial dysmorphism, cognitive impairment, pre- and post-natal growth delay, and multi-organ involvement and the clinical manifestations appear milder in cases with SMC3 and SMC1A mutations, compared with NIPBL mutations.48 Hematopoietic alterations have not been reported with cohesinopathy, nor has the development of AML. Likewise, the accumulation of aneuploidies and other chromosomal aberrations has been a recurrent feature of cohesinopathy, whereas this phenotype is largely absent in cohesin mutated AML cases, which typically present with normal karyotypes. Intriguingly, copy number gains of STAG2 or SMC1A also have been associated with cohesinopathy phenotypes,49,50,51 suggesting that there may be a critical window of adequate cohesin activity and that alterations in either direction may be detrimental. In contrast to these human data, in our mouse model, germline heterozygous Smc3 deletion was tolerated using X-linked CMV-Cre, which is expressed during early embryogenesis. The heterozygous Smc3+/−/CMV-Cre+/− female progenies had no obvious developmental defects and had normal hematopoietic homeostasis in the bone marrow (Figure 2.10). The normal hematopoietic cell numbers and differentials in the mice reflect the maintained hematopoiesis of cohesinopathies, whereas the normal number of Smc3 haploinsufficient pups contrasts with the
near absence of cohesin mutations in the human population data. This discrepancy may be due to differences between mouse and human biology; alternatively, cohesinopathy mutations may be associated with gain of function activity not recapitulated with this allele, or activity not related directly to the SMC1A/SMC3 complex.

In summary, we did not observe evidence of impaired differentiation or augmented self-renewal \textit{ex vivo} or \textit{in vivo} when \textit{Smc3} haploinsufficiency was generated using \textit{CMV-Cre}, \textit{ERT2-Cre}, and \textit{Vav1-Cre}. Instead, \textit{Smc3} haploinsufficiency was associated with competitive disadvantage, with an early bias towards phenotypes in the myeloid compartment. In AML patients, \textit{SMC3} mutations are typically early, but not initiating, genetic events. These data also suggest that pre-existing mutations may be required to enable leukemogenic consequences of \textit{SMC3} mutagenesis and to permit productive clonal expansion. Future studies are needed to determine the combination of cooperating mutations that predispose HSPCs to \textit{SMC3}-induced leukemic transformation and clonal dominance.
Figure Legends

Figure 2.1. Generation of Smc3 conditional deficient mice and allele validation.

(A) Smc3 haploinsufficient mouse model (Smc3\textsuperscript{trap/+}) was obtained from the European Mouse Mutagenesis Program (EUCOMM). Smc3 conditionally deficient mice were generated by removing the gene-trap cassette, which retains the loxP sites flanking exon 4 (Smc3\textsuperscript{fl/+}) and crossing these mice with either Vav1-Cre\textsuperscript{+/−} or ERT2-Cre\textsuperscript{+/−} to delete the allele (Smc3\textsuperscript{Δ/+}). All mice are on the C57BL/6J background. (B) Whole genome sequencing validation of Smc3\textsuperscript{fl} integration sites. (C) RNA-Seq data of the Smc3\textsuperscript{fl/+}/Vav1-Cre\textsuperscript{+/−} mice showed 227 transcripts spliced from exon 3 to 4 and then 313 transcripts from exon 4 to 5 while 279 transcripts from the other allele spliced from exon 3 to 5 (average data from 3 mice). (D) Smc3 haploinsufficiency was confirmed by reduced Smc3 level in the bone marrow (BM) cells of the Smc3\textsuperscript{fl/+}/Vav1-Cre\textsuperscript{+/−} mice measured using intracellular flow cytometry (n=5). *Denotes statistical significance by t-test. ***p<0.001. (E) Smc3 level is significantly higher in KLS (Lin-cKit+Sca1+) cells and progenitor populations than Lin- and SLAM (Lin-cKit+Sca1+CD48−CD150+). *Denotes statistical significance by one-way ANOVA with Turkey’s multiple comparisons test. ****p<0.0001.

Figure 2.2. Embryonic hematopoietic Smc3 deletion.

(A) No Smc3\textsuperscript{fl/+}/Vav1-Cre\textsuperscript{+/−} pups were observed following Smc3\textsuperscript{fl/+} and Smc3\textsuperscript{fl/+}/Vav1-Cre\textsuperscript{+/−} intercrosses (n=11 litters). *Denotes statistical significance by Chi-square test. ****p<0.0001. The E13.5 Smc3\textsuperscript{fl/+}/Vav1-Cre\textsuperscript{+/−} embryos (B) lacked gross fetal livers but retain (C) normal body weight compared with littermates. (D) The E13.5 Smc3\textsuperscript{fl/+}/Vav1-Cre\textsuperscript{+/−} embryos had decreased total fetal liver cells and (E)fetal liver hematopoietic cells (CD45.2+). (F) Myeloid
(Gr1+CD11b+) cells were reduced and increased proportions of B220+ and CD3e+ lymphocytes were observed in E13.5 $Smc3^{fl/fl}/Nav1-Cre^{+/-}$ fetal livers compared to littermate controls. (C-F) n=7 embryos per group. *Denotes statistical significance by one-way (D-E) and two-way (F) ANOVA with Turkey’s multiple comparisons test. **$p<0.01$, ****$p<0.0001$.

**Figure 2.3. Homozygous somatic $Smc3$ deletion.**

(A) $Smc3^{fl/fl}/ERT2-Cre^{+/-}$ and $Smc3^{fl/fl}$ littermate control mice were treated with 4 doses of tamoxifen (3 mg orally on days 1st, 3rd, 5th, 8th and analyzed on day 8, n = 4 mice in each group) (A) Peripheral blood analysis. (B) Body weight and spleen weight. (C) Total number of cells in the bone marrow, spleen, and thymus. (D) Analysis of lineage percentages within total bone marrow cells. (A-D) n=4 mice per group, *Denotes statistical significance by t-test. *$p<0.05$, **$p<0.01$, ***$p<0.001$. (E) Experimental schema of the $Smc3^{fl/fl}/ERT2-Cre^{+/-}$ competitive transplantation. (F) Recipient mice were treated with tamoxifen after 6-week engraftment. After tamoxifen-mediated deletion, $Smc3$ deficient cells were rapidly outcompeted, with earliest cell loss in the Gr1+ myeloid compartment, showing as complete competitive disadvantage. *Denotes statistical significance by 2-way ANOVA with Turkey’s multiple comparisons test. ****$p<0.0001$. (E-F) n=10 mice per group.

**Figure 2.4. Hematopoietic $Smc3$ haploinsufficiency.**

(A and B) Distribution of bone marrow stem, progenitor, and lineage populations in $Smc3^{fl/+}/ERT2-Cre^{+/-}$ and littermate $Smc3^{fl/+}$ mice following 9 doses of tamoxifen (n=6 mice per group). (C) Experimental schema of serial replating assay. (D-E) Colony numbers and average cells per colony on indicated week of plating in methylcellulose (n=4 mice per group). *Denotes statistical significance by t-test. *$p<0.05$, **$p<0.01$. (F) Expression analysis by RNA-Seq data
of KL (Lin-cKit+Sca1-) bone marrow cells from Smc3fl/+Vav1-Cre+/- compared to wild-type cells (n=3 mice per group). (G) Comparison of relative peak intensity identified by ATAC-Seq of KL bone marrow cells from relative peak Smc3fl/+Vav1-Cre+/- compared to wild-type cells (n=3 mice per group).

**Figure 2.5. Competitive transplantation of Smc3 haploinsufficient bone marrow cells.**

(A-F) Competitive repopulation assay using Smc3fl/+ERT2-Cre+/- BM cells and littermate Smc3fl/+ BM cells with competitor CD45.1 x CD45.2 bone marrow cells (3 donor mice per group and 10 recipient mice per group). Following 6 weeks of engraftment, equal peripheral chimerism was validated and recipient mice were treated with 9 doses of tamoxifen. (B - C) Following 42 weeks, bone marrow chimerism was analyzed (n=3 mice per group). *Denotes statistical significance by t-test, *p<0.05, **p<0.01, ****p<0.0001. (D-F) At interval time-points during follow-up peripheral blood chimerism was evaluated within the Gr1, B220, and CD3e compartments. *Denotes statistical significance by 2-way ANOVA with Turkey’s multiple comparisons test, *p<0.05, **p<0.01, ****p<0.0001. (G-J) Competitive repopulation assay of Smc3fl/+ERT2-Cre+/- BM cells under hematopoietic stresses (n=10). As before, recipient mice were treated with 9 doses of tamoxifen after 6-week engraftment. PipC and 5-FU were given 16 weeks post-transplant, respectively. *Denotes statistical significance by 2-way ANOVA with Turkey’s multiple comparisons test, **p<0.01, ***p<0.001, ****p<0.0001.

**Figure 2.6. Effect of Dnmt3a haploinsufficiency on competitive disadvantage in Smc3 haploinsufficient BM cells.**

(A-D) Competitive repopulation assay of Smc3fl/+ERT2-Cre+/-/Dnmt3a+/- BM cells and indicated littermate controls (n=10 mice per group). As in Figure 2.5, total bone marrow cells were
allowed to engraft for 6 weeks and equivalent chimerism was validated before treatment of all cohorts with 9 doses of tamoxifen (3 mg/day). Peripheral blood chimerism was evaluated by flow cytometry at indicated time points. (E) Bone marrow chimerism assessed by flow cytometry 26 weeks after engraftment. KL: Lin-cKit+Sca1-. (n=4 mice in each group). *Denotes statistical significance by 2-way ANOVA with Turkey’s multiple comparisons test. *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001.

Figure 2.7. Splicing analysis of exon 3 to exon 5 in wild-type and Smc3fl/fl/Vav1-Cre+/- KL cells.

Lin-cKit+Sca1- bone marrow cells were subjected to RNA-Seq (Figure 2.1C and 2.4F). Schema indicates total number of reads spanning each splice junction from indicated mice.

Figure 2.8. Representative plot of intracellular flow cytometry data (Figure 2.1D-E).

(A) Percentages of Smc3+ cells (left) and mean fluorescence intensity (MFI) (right) of WBM from mice used in Figure 2.1D. (B). Overlay of the two MFI plots. (C) Percentages of Smc3+ cells (left) and mean fluorescence intensity (MFI) (right) of lin-, KLS, SLAM, and KL cells from mouse used in Figure 2.1E. The height of the peak is proportional to the number of events collected.

Figure 2.9. Analysis of homozygous somatic Smc3 deletion.

(A) Western blot of Smc3 in total bone marrow cells following 4 doses of tamoxifen in indicated mice. (B) Image of spleens from Smc3fl/fl/ERT2-Cre+/- and Smc3fl/fl littermate controls following 4 doses of tamoxifen. (C-D) Proportion of bone marrow cells and thymocytes with indicated immunophenotypes following 4 doses of tamoxifen in Smc3fl/fl/ERT2-Cre+/- mice and littermate
controls (n=4 mice per group), *Denotes statistical significance by t test, *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001.

**Figure 2.10. Analysis of germline heterozygous Smc3 deletion.**

(A) Numbers of male vs. female pups generated from Smc3^{fl/} and CMV-Cre^{+/} intercrosses (of note, CMV-Cre is X-linked). (B-E) Complete blood counts, total number of bone marrow cells, percentages of HSPCs, and cells of myeloid (Gr1+ CD11b+), B cells (B220+ CD4-), and erythroid cells (Ter119+ CD71+) in Smc3^{+/}/CMV-Cre^{+/} females and Smc3^{fl/+} littermate males, (n=4 mice per group).

**Figure 2.11. Immunophenotypic analysis of colonies in serial replating assay ex vivo (Figure 2.4C-E).**

(A) Percentages of Gr1+ CD11b+ and cKit+ FcεR1+ cells in the Smc3^{fl/+} and Smc3^{fl/+}/ERT2-Cre^{+/} colonies week 1-4 respectively (n=4 mice per group). (B) Representative plot of the Smc3^{fl/+} and Smc3^{fl/+}/ERT2-Cre^{+/} colonies week 1-4. *Denotes statistical significance by t test, *p<0.05.

**Figure 2.12. Competitive transplantation of ERT2-Cre^{+/} bone marrow cells.**

(A) Competitive repopulation assay using ERT2-Cre^{+/} BM cells with competitor CD45.1 x CD45.2 bone marrow cells (3 donor mice and 5 recipient mice). Following 6 weeks of engraftment, equal peripheral chimerism was validated and recipient mice were treated with 9 doses of tamoxifen. (B) At interval time-points during follow-up peripheral blood chimerism was evaluated within the Gr1, B220, and CD3e compartments.
Figure 2.1. Generation of Smc3 conditional deficient mice and allele validation.

A.

B.

Whole genome sequencing

C.

RNA-Seq

Smc3<sup>fl</sup>/Vav1-Cre<sup>+</sup>

Ave Reads: 227  Ave Reads: 313  Ave Reads: 279

D.

E.

Smc3 MFI of bone marrow cells

Smc3<sup>fl</sup>  Smc3<sup>fl</sup>/Vav1-Cre<sup>+</sup>
Figure 2.2. Embryonic hematopoietic Smc3 deletion.

A. Number of viable pups born

B. Smc3<sup>3/4</sup> Smc3<sup>3/4</sup>/Vav1-Cre<sup>−/−</sup> Smc3<sup>3/4</sup>/Vav1-Cre<sup>−/−</sup>

C. E13.5 embryo body weight (gram)

D. Fetal liver cells per E13.5 embryo

E. %CD45<sup>+</sup> cells in E13.5 fetal liver

F. % of total E13.5 fetal liver cells

---

68
Figure 2.3. Homozygous somatic Smc3 deletion.

A. Hematopoietic parameter after 4 doses of tamoxifen.

B. Weights (grams) after 4 doses of tamoxifen:
- Body weight
- Spleen weight

C. Total number of cells after 4 doses of tamoxifen:
- Bone marrow
- Spleen
- Thymus

D. % of cells in bone marrow after 4 doses of tamoxifen:
- cKit+
- Gr1+
- CD11b+
- B220+
- CD8+
- CD4+
- cKit+ Ter119
- CD71+
- CD71+

E. Cohort 1: Smc3<sup>fl/fl</sup>  
Cohort 2: Smc3<sup>fl/fl</sup>/ERT2-Cre<sup>Cre<sup>+</sup></sup>

F. %CD45.2 chimerism in peripheral blood:
- Weeks following transplant
Figure 2.4. Hematopoietic Smc3 haploinsufficiency.

A. 

B. 

C. 

D. 

E. 

F. 

G. 

Samples with ave fpkms in one condition >1
Figure 2.5. Competitive transplantation of Smc3 haploinsufficient bone marrow cells.

A. 

B.

C.

D.

E.

F.

G.

H.

I.

J.

9 doses of tamoxifen
Figure 2.6. Effect of Dnmt3a haploinsufficiency on competitive disadvantage in Smc3 haploinsufficient BM cells.

A. 

B. 

C. 

D. 

E. 

- Smc3fl/+ 
- Smc3fl/+ERT2-Cre0/-. 
- Smc3fl/+Dnmt3a0/-. 
- Smc3fl/+ERT2-Cre0/-.Dnmt3a0/- 

9 doses of tamoxifen

9 doses of tamoxifen

n.s.
Figure 2.7. Splicing analysis of exon 3 to exon 5 in wild-type and Smc3^fl/fl/Vav1-Cre^+/- KL cells.

A. Mouse 10433, Wild type

B. Mouse 10410, Smc3^fl/fl/Vav1-Cre^+/-

C. Mouse 10442, Wild type

D. Mouse 10411, Smc3^fl/fl/Vav1-Cre^+/-

E. Mouse 10841, Wild type

F. Mouse 10412, Smc3^fl/fl/Vav1-Cre^+/-
Figure 2.8. Representative plot of intracellular flow cytometry data (Figure 2.1.D-E).

A. Mouse 8012, Smc3<sup>fl</sup>, WBM

Mouse 8013, Smc3<sup>fl</sup>/Vav1-Cre<sup>+</sup>, WBM

C.

Mouse 10726, Wild type, lin-

Mouse 10726, Wild type, KLS

Mouse 10726, Wild type, SLAM

Mouse 10726, Wild type, KL

Smc3-APC
Figure 2.9. Analysis of homozygous somatic Smc3 deletion.

A.

B.

C.

D.
Figure 2.10. Analysis of germline heterozygous Smc3 deletion.

A. 

B. 

C. 

- **Smc3<sup>+/−</sup>** Male
- **Smc3<sup>+/−</sup>/CMV-Cre<sup>+/−</sup>** Female

D. 

E. 

- **Smc3<sup>+/−</sup>** Male
- **Smc3<sup>+/−</sup>/CMV-Cre<sup>+/−</sup>** Female
Figure 2.11. Immunophenotypic analysis of colonies in serial replating assay *ex vivo* (Figure 2.4.C-E).

A.  

B. 

<table>
<thead>
<tr>
<th>Week</th>
<th>Smc&lt;sup&gt;3&lt;no-break/&gt;&lt;/sup&gt;</th>
<th>Smc&lt;sup&gt;3&lt;no-break/&gt;-ERT2-Cre&lt;sup&gt;1&lt;no-break/&gt;&lt;/sup&gt;/</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td><img src="image1" alt="Image" /></td>
<td><img src="image2" alt="Image" /></td>
</tr>
<tr>
<td>2</td>
<td><img src="image3" alt="Image" /></td>
<td><img src="image4" alt="Image" /></td>
</tr>
<tr>
<td>3</td>
<td><img src="image5" alt="Image" /></td>
<td><img src="image6" alt="Image" /></td>
</tr>
<tr>
<td>4</td>
<td><img src="image7" alt="Image" /></td>
<td><img src="image8" alt="Image" /></td>
</tr>
</tbody>
</table>

* Percent of cells in retrieved methylcellulose colonies
Figure 2.12. Competitive transplantation of $ERT2-Cre^{+/−}$ bone marrow cells.
References


Chapter 3:
Exome analysis of treatment-related AML after APL suggests secondary evolution
Treatment-related acute myeloid leukemia (tAML) and treatment-related myelodysplastic syndrome (tMDS) have been associated with many types of chemotherapy and radiation.\(^1\)

Treatment-related AML or tMDS have been observed after treatment of acute promyelocytic leukemia (APL) with combination all-trans retinoic acid (ATRA)/idarubicin/cytarabine.\(^2,3,4,5,6\)

It is unknown whether tAML will emerge following exposure to all-trans retinoic acid (ATRA) and arsenic trioxide (ATO), which should not cause DNA damage or the clonal selection of chemotherapy-resistant clones that give rise to tAML.\(^1,6,7\)

We describe a case of an older woman who presented initially with dysplasia. Shortly thereafter she developed APL and was treated with ATRA/ATO. Five years later she developed what clinically appeared to be tAML. Exome sequencing revealed a founding clone with a \textit{TET2} mutation and shared passenger mutations that existed at all three time points. The APL sample shared these mutations, and the subsequent “tAML” emerged as a new subclone with an \textit{NPM1} mutation, more consistent of secondary AML rather than tAML. This case demonstrates that APL may emerge within the context of clonal hematopoiesis, and that tAML emerging after ATRA/ATO should be evaluated for features consistent with secondary AML.

An 81 year-old woman presented initially with cytopenias at an outside facility. A bone marrow biopsy was performed, revealing trilineage dysplasia, 6% promyelocytes, and a normal karyotype, 46 XX[20/20] (Table 3.1). Two months later, the cytopenias persisted and a repeat bone marrow aspirate was hypercellular with 50% promyelocytes. Cytogenetics revealed 46 XX, t(15;17)[18]/46 XX[2] and 89% of cells were positive for \textit{PML-RARA} by FISH. She was referred to Washington University. The bone marrow biopsy was repeated and banked with appropriate consent for genomic analysis. The repeat biopsy was unfortunately hemodilute with 4.5% t(15;17) by FISH [9/200], was not evaluated by cytomorphology, and RT-PCR failed (control
GAPDH primers did not amplify). She was treated with ATRA/ATO. Bone marrow biopsy 6 weeks later revealed 1% promyelocytes and normal karyotype by cytogenetics and FISH. Due to her age, no further bone marrow biopsies were performed and subsequent peripheral blood RT-PCR tests were negative. Five years later she became increasingly cytopenic and a bone marrow aspirate revealed acute myeloid leukemia with 72% blasts, 20% promyelocytes, and a normal karyotype. She was treated with decitabine. She deteriorated during the first cycle and transitioned to hospice.

Exome sequencing was performed on three samples using techniques described elsewhere and compared against a skin sample, which was used as a germline control (Figure 3.1). Cells retained on a coverslip were available from the dysplastic pre-APL sample and this was used for genomic analysis. No coverslip or other material was available from the APL sample with 89% PML-RARA. Cryopreserved bone marrow aspirate cells were used for APL (4.5% PML-RARA) and AML (72% blasts) analysis. The collected APL sample only generated one cryovial, which was used for DNA synthesis. Unfortunately, no additional samples are available for RNA-Seq analysis or sequencing of subpopulations after flow sorting.

A shared founding clone existed across all three time points that contained a TET2 mutation and 15 additional variants (Figure 3.1, black). Three additional clusters were identified: 1) variants that were present predominantly in the dysplasia sample (SEMA4A, and ZBTB7A, orange); 2) variants that were absent in the AML sample (TET1, SACM1L, OR7D2, SH3TC1, blue); 3) variants that increased in the AML sample, but were present at low variant allele frequencies or were undetected in prior samples (NPM1 and 13 additional variants, red).

Treatment-related AML has been associated with APL therapy that includes alkylating agents, but has not been associated with ATRA/ATO, which are non-cytotoxic and do not damage
DNA. This unusual case initially appeared consistent with tAML following ATRA/ATO, however, following exome analysis, appears to be more consistent with a pre-existing dominant clone associated with dysplasia and a TET2 mutation, and an NPM1-associated secondary AML. Two models are possible (Figure 3.1F). First, the APL may have emerged as a subclone of the TET2 founding clone. Given the high TET2 variant allele frequency (VAF, consistent with 70% and 66% tumor burden in the dysplastic and APL samples, respectively) and the high tumor burden of PML-RARA in the bone marrow (85%), this model seems likely. Alternatively, because the sequenced APL sample is hemodilute, it is possible that the PML-RARA clone is independent of the TET2 clone, that the APL clone did not peripheralize, or that geographical heterogeneity existed in this older patient. Given the absence of additional samples, it is impossible to determine whether the APL emerged as a subclone of the TET2 founding clone occurred independent of the TET2 clone.

In order to determine if these subclonal patterns are observed in other APL patients, we reviewed available cytogenetic results from published APL patients. Two cases of disease progression from MDS to APL have been described10, and cases of treatment-related APL have been described following MDS.11 In the former cases, like this case, the timeline of evolution and dysplastic changes in the APL morphology suggests that the APL clone likely was evolutionarily related to the MDS clone. In the latter cases, the cytogenetics and timeline suggest the APL clone likely emerged independent of the MDS clone.

Clonal hematopoiesis of indeterminate potential (CHIP) and MDS have been associated with deletions involving chromosomes 5 and 712,13, which occasionally co-occur with t(15;17).14,15,16 Therefore, we examined the subclonal relationship of t(15;17) with possible CHIP-related variants among APL patients enrolled at large cancer centers. We identified 44 APL cases with
t(15;17) and cytogenetic abnormalities in chromosomes 5 or 7. Five cases involved chromosome 5, two of which also had abnormalities in chromosome 7. Thirty-three cases involved a monoclonal process, suggesting that the variants in chromosomes 5 or 7 co-occurred with t(15;17). Of the eleven cases with identified polyclones, the chromosome 5 or 7 variant was in a subclone of t(15;17) in ten cases and in only one case was there evidence of del(7) in a clone that was independent of the t(15;17) clone (Patient 52009 from APL 2006, 46, XY, t(15,17)(q22, q11)[13]/46, XY, del(7)(q35)[5]/46, XY [2]) (Table 3.2). These data suggest that t(15;17) may co-occur with additional cytogenetic deletions of chromosomes 5 and 7, but t(15;17) is almost invariably the founding event, and it is unusual for a concurrent CHIP or MDS clone to co-exist with t(15;17).

Other investigators have reviewed the outcomes in APL patients with cytogenetic abnormalities that occur in addition to t(15;17).\textsuperscript{16,17,18} The most recurrently observed co-occurring cytogenetic abnormalities include +8 and +21, both of which frequently can be identified as subclonal progression events when analyzing metaphase cytogenetics or FISH, and have been observed as progression events in MDS and AML.\textsuperscript{11} In these studies, additional cytogenetic abnormalities have not correlated with initial clinical characteristics, or with outcomes in patients with t(15;17).\textsuperscript{16,17,18}

Morphologic disease switching also has been described in NPM1-mutated AML patients who subsequently developed MDS or myelofibrosis. In each of these cases, the NPM1-mutation was lost, and the MDS or myelofibrosis evolved from an antecedent clone with a mutation in TET2, JAK2, ASXL1, IDH2, or a spliceosome transcript, suggesting that the two clonal diseases were related through an ancestral clone.\textsuperscript{19}
In summary, we present a case of what initially appeared to be tAML following non-cytotoxic ATRA/ATO therapy for APL. Exome analysis clearly demonstrated a TET2-associated, dominant clonal process that anteceded the APL diagnosis, persisted, and gave rise to AML associated with evolutionary expansion of an NPM1-mutated subclone. This progression would be more consistent with a secondary AML process, rather than a treatment-related process. Additional characterization of this case would be interesting. Unfortunately, samples for such analyses are unavailable. Review of additional cytogenetic abnormalities observed in APL patients did identify one rare case with del(7) independent of the APL clone, suggesting that APL can co-exist with CHIP or MDS clones, although it appears much more common for such cytogenetic abnormalities to occur as subclones of the APL clone. Collectively, these results suggest that caution must be exercised when interpreting the development of tAML following ATRA/ATO therapy, and subclonal expansion of related or well-established clones should be considered, especially in older patients.
Figure Legends

Figure 3.1. Exome analysis of patient 10DD-1029.

(A) Summary results of all somatic variants detected at any of the three time points analyzed. (B) Cluster of variants with stable VAFs across all three samples. (C) Cluster of variants present in the initial dysplastic sample, with reduced VAFs in subsequent samples. (D) Cluster of variants with VAFs in the initial two samples, but which were absent in the subsequent AML sample. (E) Cluster of variants associated with the AML progression, which were largely absent in the initial two samples. (F) Two models for subclonal expansion.
Figure 3.1. Exome analysis of patient 10DD-1029.
### Tables 3.1. Clinical data of the patient.

<table>
<thead>
<tr>
<th></th>
<th>Dysplasia</th>
<th>APL</th>
<th>APL (banked)</th>
<th>post-APL</th>
<th>AML</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Day 0</td>
<td>Day 49</td>
<td>Day 51</td>
<td>Day 87</td>
<td>Day 1897</td>
</tr>
<tr>
<td>Sequencing</td>
<td>exome</td>
<td>ND</td>
<td>Exome</td>
<td>ND</td>
<td>exome</td>
</tr>
<tr>
<td>t(15;17)</td>
<td>0%</td>
<td>89%</td>
<td>4.5%</td>
<td>0%</td>
<td>0%</td>
</tr>
<tr>
<td>BM blasts</td>
<td>0%</td>
<td>1%</td>
<td>ND</td>
<td>1%</td>
<td>72%</td>
</tr>
<tr>
<td>BM promyelocytes</td>
<td>6%</td>
<td>50%</td>
<td>ND</td>
<td>1%</td>
<td>20%</td>
</tr>
<tr>
<td>BM myelocytes</td>
<td>9%</td>
<td>25%</td>
<td>ND</td>
<td>25%</td>
<td>1%</td>
</tr>
<tr>
<td>BM metamyelocytes</td>
<td>2%</td>
<td>3%</td>
<td>ND</td>
<td>3%</td>
<td>5%</td>
</tr>
<tr>
<td>BM bands</td>
<td>9%</td>
<td>2%</td>
<td>ND</td>
<td>2%</td>
<td>0%</td>
</tr>
<tr>
<td>BM dysplasia</td>
<td>trilineage</td>
<td>atypia</td>
<td>ND</td>
<td>none</td>
<td>none</td>
</tr>
<tr>
<td>PB WBC</td>
<td>NA</td>
<td>0.9</td>
<td>0.9</td>
<td>0.9</td>
<td>2.2</td>
</tr>
<tr>
<td>PB % Lymphs</td>
<td>NA</td>
<td>95%</td>
<td>95%</td>
<td>95%</td>
<td>88%</td>
</tr>
<tr>
<td>PB % Blasts</td>
<td>NA</td>
<td>0%</td>
<td>0%</td>
<td>0%</td>
<td>6%</td>
</tr>
</tbody>
</table>

AML, acute myeloid leukemia; APL, acute promyelocytic leukemia; BM, bone marrow; NA, not available; ND, not done; PB, peripheral blood; WBC, white blood cells.
Table 3.2. APL patients with cytogenetic abnormalities in chromosomes 5 or 7 and evaluable subclonal architecture.

<table>
<thead>
<tr>
<th>Source</th>
<th>Patient</th>
<th>Karyotype</th>
<th>t(15;17) subclone</th>
<th>Clone independent of t(15;17)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MD Anderson</td>
<td>NA</td>
<td>46,XX,add(7)(q32),t(15;17)(q24;q21)[5] /47,idem,+8[2]/46,XX,del(7)(q22q34),t(15;17)[7]/46,XX,add(4)(p16),t(15;17)[5]/46,XX[1]</td>
<td>del(7), +8, add(4)</td>
<td></td>
</tr>
<tr>
<td>APL 2006</td>
<td>78034</td>
<td>t(15 ;17), +8 [ ?], t(15 ;17), -7, -5 [ ?]</td>
<td>-7, -5, +8</td>
<td></td>
</tr>
<tr>
<td>APL92</td>
<td>108</td>
<td>46,XX,1p-,7q-,15q+,17q-,19q+ [14/20] / 46,XX,15q+,17q- [6/20]</td>
<td>del(1p), del(7q), add(19q)</td>
<td></td>
</tr>
<tr>
<td>APL92</td>
<td>687</td>
<td>46,XX,t(15;17)(q22;q11 ~ 21) [19/20] / 46,idem,-7,+mar [1/20]</td>
<td>-7</td>
<td></td>
</tr>
<tr>
<td>APL92</td>
<td>742</td>
<td>46,XX,t(15;17) [16/17] / 46,idem,add(7)(q?) [1/17]</td>
<td>add(7)</td>
<td></td>
</tr>
<tr>
<td>MD Anderson</td>
<td>NA</td>
<td>46,XX,t(15;17)(q24;q21)[15]/46,idem,del(7)(q22q32)[3]; 46,XX[2]</td>
<td>del(7)</td>
<td></td>
</tr>
<tr>
<td>MD Anderson</td>
<td>NA</td>
<td>46XX,t(15;17)(q22;q21.1)[16]/46,idem,del(7)(q22q34)[1]/ 46XX[3]</td>
<td>del(7)</td>
<td></td>
</tr>
<tr>
<td>MD Anderson</td>
<td>NA</td>
<td>46XY,t(15;17)(q22;q21)[16]/46,idem,del(7)(q32q36)[1]/46,XY[3]</td>
<td>del(7)</td>
<td></td>
</tr>
<tr>
<td>MD Anderson</td>
<td>NA</td>
<td>46XX, del(7)(q31q36), inv(9)(p11;q12),t(15;17)(q22;q21)[2]/47XX,+8,inv(9), der(15), der(17)[3]/46XX, inv(9).</td>
<td>del(7), +8</td>
<td></td>
</tr>
</tbody>
</table>
References


11. Andersen MK, Larson RA, Mauritzson N, Schnittger S, Jhanwar SC, Pedersen-Bjergaard J. Balanced chromosome abnormalities inv(16) and t(15;17) in therapy-related


