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Structured growth and genetic drift raise relatedness in the social amoeba Dictyostelium discoideum

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1. Introduction

 The fitness and success of a group of cooperators is under constant threat of collapse by the infiltration and spread of selfish individuals, known as cheaters, that reap the benefits of cooperation without paying the costs (1, 2). Kin selection theory predicts that cooperative behaviours will be selected if relatives preferentially interact with one another by kin recognition or high population viscosity (3-5). High relatedness is key to kin selection and has been shown in the social insects (6, 7), birds (8, 9), mammals (10, 11) and microbes (12-14). When a microbial colony grows from a single point, zones of highly-related individuals can form when genotypes, identical except for their fluorescent labels, are mixed in moderate numbers (~1000)(15). Models (16) suggest that the mechanism behind this is genetic drift; only a few cells are actively dividing in any region of the growing colony edge and there are no phenotypic differences between cells (**Fig 1A**). Selection could also produce zones, but it is not required. Simulations suggest that segregation rate is dependent upon availability of nutrients; diffusible nutrients cannot penetrate far into the colony, lowering the effective population and promoting segregation by drift. This outward growth results in the formation of growing zones of clonemates (**Fig 1A**). Further simulations show that when genotypes have clear altruist or cheater phenotypes, altruists obtain the group benefit of their behaviour because of increased relatedness through structured growth (16). However, the effect structured growth has on relatedness has not been quantitatively assessed for any social microbe, and so to address this issue, we use the social amoeba

scarce, starving *Dictyostelium* cells aggregate to form a fruiting body where around one-fifth die to form

a stalk, lifting the remaining cells aloft as viable spores (20). Such a costly act of altruism must have

Dictyostelium discoideum, now a model organism for social evolution studies (17-19). When food is

 evolved in groups of highly-related individuals (3, 4, 21). However, during the aggregation stage, several genotypes mix together forming chimeras (22). Cheater genotypes avoid contributing their fair share of cells to the stalk and are overrepresented in the sporehead (18, 22).

 We investigate whether genetic drift during the non-social growth stage can act as a mechanism for the formation of highly-related groups prior to the social stage. Whether it will form clear zones like bacteria is not obvious given the high mobility of *Dictyostelium* amoebae. Natural formation of zones of high relatedness during growth should increase the opportunity for altruism to evolve and be maintained, even in initially mixed populations (**Fig 1**).

 Relatedness within fruiting bodies should increase with distance from the origin because the zones widen with distance whilst the area of an aggregation territory for a given density of amoebae remains constant, increasing the likelihood that cells of only one genotype will fall within an aggregation territory (23) (**Fig 1B**).

 Here, we demonstrate that even with large initial populations and mobile cells, founder effects and genetic drift at the growing edge act as a mechanism to increase and maintain relatedness in *D. discoideum* and that this effect is greater in low food conditions. We also show that clear zones can form from a single cell.

2. Material and Methods

 Wild *D. discoideum* clones were grown in the presence *of Klebsiella aerogenes* (Ka) bacteria on 62 SM agar (10g peptone, 1g yeast extract, 10g glucose, 1.9g KH₂PO₄, 1.3g K₂HPO₄, 0.49g MgSO₄ (anhydrous), 17g agar per litre) plates and incubated 22°C. We obtained amoebae by spreading spores evenly over SM plates in the presence of Ka. Log-phase cells were harvested and washed by repeated 65 centrifugation in KK2 buffer (16.1mM KH₂PO₄, 3.7mM K₂HPO₄).

 Clones were transformed with actin15-RFP following the procedure in Pang et al. (24). Clones expressing the markers were then further selected on G418-SM agar plates (30µg G418/ml). Mixes were performed with two clones that were isogenic except for the RFP marker, to eliminate any effects of kin recognition.

Genetic drift assay

 Log-phase *NC28.1* and *NC28.1*-RFP cells were mixed in three proportions (wt:RFP; 50:50, 95:5 72 and 99:1) and suspended at a density of 4×10^8 cells/ml in KK2 with 1×10^7 total cells pipetted per plate in the centre of a 150mm wide SM plate spread with Ka. The low-nutrient treatment diluted nutrients 50- fold. Images of whole plates were taken using an SLR digital camera with a 660 nm filter and a 514nm LED as a light source (25).

 Ten fruiting bodies were taken at each of four distances (at <1, 3, 5 and 7 cm) from the centre and suspended in spore buffer (20mM EDTA and 0.1% NP-40). The proportion of the two genotypes in each fruiting body was then determined by direct counting of spores using a fluorescent microscope. The relatedness (*r*) within individual sporeheads was calculated as:

$$
r = \frac{p(p - \bar{p})}{(1 - \bar{p})} + \frac{q(q - \bar{q})}{(1 - \bar{q})}
$$

 where *p* and *q* are the proportion of the two genotypes in the sporehead and are the proportions of *p* and *q* for the whole population (26).

3. Results

 For all three mixtures of NC28.1 and NC.28-RFP, clear zones formed readily (**Fig 2A**), and as a consequence of structured growth, relatedness within fruiting bodies increased with distance from the 86 origin, where relatedness was zero $(R^2 = 0.5944; P = 0.0033;$ Fig 2B). Clones did mix; there was low relatedness in fruiting bodies at the 'borders' between zones.

- A fortuitous mutation showed that a clonal zone can form from a single mutant cell. On a clonal plate of the genotype *NC34.1* a spontaneous mutation occurred during growth producing a zone of cells that could not form normal fruiting bodies (**Fig 2C**).
- When the availability of food was decreased, relatedness almost doubled, reaching 0.68 at the 92 plate edge, compared to 0.36 with the regular SM plate $(R^2 = 0.7576, P < 0.001; 2$ -way ANOVA: nutrient level, *F*1,24 = 12.7, *P* = 0.003; distance from origin, *F*3,24 = 18.1, *P* < 0.001; **Fig 2B**).
- For data see the supplemental material.

4. Discussion

 In any given population, the fitness of an altruistic gene is dependent upon the structure of the population (3, 4, 27, 28). Our results show that high relatedness can be obtained by structured growth and genetic drift without a small initial group size (**Fig 2A**). This result is important for two reasons. First, high relatedness means there will be distinct local groups of cooperators and cheaters, so that the cooperators are protected from exploitation. Second, these results provide one explanation for the disparity between ready chimera formation *in vitro* (22, 29), and high relatedness in wild fruiting bodies (19).

 This, however, provides another problem; if local dispersal produces areas of high relatedness to neighbours, then it also makes neighbours closest competitors, cancelling out the relatedness effects (5). However, combining structured growth with regular spore dispersal can maintain altruism, as spore dispersal relaxes the effects of competing with highly-related neighbours (5).

 During the evolution of multicellularity a degree of unrelatedness – and therefore conflict – had to be overcome, perhaps by recognition or via some other mechanism e.g. (30-33). We show that a sufficient degree of structured growth may be sufficient to achieve this, despite the mobility of amoebae that tends to mix cells. Relatedness also increased at a higher rate when there were fewer nutrients, creating more distinct zones (**Fig 2B**). This suggests that in the wild where there are likely to be even fewer nutrients, high relatedness may be generated even more readily. Thus, combined with the added friction of a rough natural substrate this effect upon relatedness may be further enhanced. Other mechanisms such as recognition (30) may have evolved to inflate relatedness further, an important adaptation if motile vegetative *Dictyostelium* cells cause mixing of genotypes slowing the rate of increase. Our study shows that structured growth is a potentially important cause of high relatedness. Once altruism evolves, groups of altruists can form whilst excluding cheaters, allowing more complex

mechanisms to evolve.

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Figures

Figure 1: Zones of high relatedness form by genetic drift. (A) A thin layer of red and green cells,

identical except for their colour actively grow at the colony edge, simple sampling error creates small

groups of the same genotype. Cells growing in the direction of the arrows form zones of coloured cells.

 (B) Relatedness increases within fruiting bodies with distance from centre of origin (O). Each aggregation territory (blue-grey lines) is of a similar size, so the probability a territory is contained within a single zone increases with distance from O.

 Figure 2: Zones of high relatedness form readily without recognition. (A) *NC28.1* mixed in three different proportions with *NC28.*1-RFP. The following mixes were (wt:RFP): (i) 50:50, (ii) 95:5 and (iii) 99:1. In each case distinct zones of *28.1*-RFP formed. (B) Relatedness (*r*) within sporeheads increases from the point of origin in a 50:50 mix of *NC28.1* and *NC28.1*-RFP. When food was plentiful (dashed line) 137 r increased significantly with distance from origin (O) $(R^2 = 0.5944, P = 0.003)$. When food was reduced 138 (solid line), *r* also increased (R^2 = 0.7576, P < 0.001) and to a significantly higher level than when grown on plentiful food (2-way ANOVA: nutrient level *F*1,24 = 12.7, *P* = 0.003; distance form origin: *F*3,24 = 18.1, *P* < 0.001). (C) A non-fruiting mutant forms a zone from a population of originally clonal cells of *NC34.1*. The mutation occurred at some point between O and the apex of the zone (dashed line). The wide angles suggest the mutant had an accelerated growth rate.

 1. West SA, Diggle SP, Buckling A, Gardner A, Griffins AS. 2007 The social lives of microbes. *Annu Rev Ecol Evol* S, **38**,53-77.

 2. West SA, Griffin AS, Gardner A, Diggle SP. 2006. Social evolution theory for microorganisms. *Nat Rev Microbiol* **4**,597-607.

3. Hamilton WD. 1964 Genetical evolution of social behaviour I. *J Theor Biol* **7**,1-16.

4. Hamilton WD. 1964 Genetical evolution of bocial behaviour 2. *J Theor Biol* **7**,17-52.

5. Queller DC. 1992 Does population viscosity promote kin selection. *Trends in Ecol Evol* **7**,322-4.

- 18. Ennis HL, Dao DN, Pukatzki SU, Kessin RH. 2000 *Dictyostelium* amoebae lacking an F-box protein
- form spares rather than stalk in chimeras with wild type. *P Natl Acad Sci* **97**,3292-7.
- 19. Gilbert OM, Foster KR, Mehdiabadi NJ, Strassmann JE, Queller DC. 2007 High relatedness
- maintains multicellular cooperation in a social amoeba by controlling cheater mutants. *P Natl*
- *Acad Sci* **104**,8913-7.
- 20. Raper KB. 1984 *The Dictyostelids*. Princeton, New Jersey: Princeton University Press.
- 21. Bourke AFG. 2011 *Principles of Social Evolution*. New York: Oxford University Press.
- 22. Strassmann JE, Zhu Y, Queller DC. 2000 Altruism and social cheating in the social amoeba *Dictyostelium discoideum*. *Nature* 408,965-7.
- 23. Bonner JT, Dodd MR. 1962 Aggregation territories in the cellular slime molds. Biol Bull **122**,13- 24.
- 24. Pang KM, Lynes MA, Knecht DA. 1999 Variables controlling the expression level of exogenous genes in *Dictyostelium*. *Plasmid* **41**,187-97.
- 25. Chin-Sang I, Zhong W. 2009 Using LEDs as a low-cost source to detect GFP and DsRED. *The Worm Breeder's Gazette* **18**,6.
- 26. Queller DC, Goodnight KF. 1989 Estimating relatedness using genetic-markers. *Evolution* **43**,258- 75.
- 27. Griffin AS, West SA, Buckling A. 2004 Cooperation and competition in pathogenic bacteria. *Nature* **430**,1024-7.
- 28. West SA, Pen I, Griffin AS. 2002 Conflict and cooperation cooperation and competition between relatives. *Science* **296**,72-5.

- 29. Buttery NJ, Rozen DE, Wolf JB, Thompson CRL. 2009 Quantification of social behavior in *D. discoideum* reveals complex fixed and facultative strategies. *Curr Biol* **19**,1373-7. 30. Benabentos R, Hirose S, Sucgang R, Curk T, Katoh M, Ostrowski EA, Strassmann JE, Queller DC, Zupon B, Shaulsky G et al. 2009 Polymorphic members of the lag gene family mediate kin discrimination in *Dictyostelium*. *Curr Biol* **19**,567-72. 203 31. Foster KR, Shaulsky G, Strassmann JE, Queller DC, Thompson CRL. 2004 Pleiotropy as a mechanism to stabilize cooperation. *Nature* **431**,693-6. 32. Ostrowski EA, Katoh M, Shaulsky G, Queller DC, Strassmann JE. 2008 Kin Discrimination increases with genetic distance in a social amoeba. *PLOS Biol* **6**,2376-82. 33. Queller DC, Ponte E, Bozzaro S, Strassmann JE. 2003 Single-gene greenbeard effects in the social
- amoeba *Dictyostelium discoideum*. *Science* **299**,105-6.
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Fig 1

